

Review
Current State of Bioceramics

S.V. Dorozhkin*

Kudrinskaja sq. 1–155, Moscow 123242, Russia

received March 23, 2018; received in revised form May 16, 2018; accepted July 13, 2018

Abstract

In the late 1960s, strong interest grew in studying various types of ceramics as potential bone grafts thanks to their suitable biomechanical properties. A bit later, such synthetic biomaterials were termed bioceramics. Since then, there has been a number of important achievements in this field. Namely, after the initial development of bioceramics that were just tolerated in the physiological environment, emphasis was shifted towards those able to form direct chemical bonds with the adjacent bones and tissues. Afterwards, based on selection of the appropriate chemical composition coupled with structural and compositional controls, it became possible to choose whether the bioceramic implants remained biologically stable once incorporated into the skeletal structure or whether they should be resorbed over time. At the turn of the millennium, a new concept of regenerative bioceramics was developed and such formulations became an integrated part of the tissue engineering approach. Now bioceramic scaffolds are designed to induce bone formation and vascularization. These scaffolds are usually porous and often harbor various biomolecules and/or cells. This review describes the major types and properties of bioceramics suitable for tissue engineering.

Keywords: Bioceramics, biomaterials, grafts, biomedical applications, tissue engineering

I. Introduction

The field of biomaterials requires the input of knowledge from very different areas of science and technology so that the implanted material performs adequately in a living body. This discipline was founded in the knowledge of materials science and biological clinical science with the final aim of achieving the correct biological interaction between the implanted material and the living body. Therefore, biomaterials appear to be an excellent example of a truly multi-disciplinary field, in which the material, developed by materials scientists and engineers, has to be validated and must perform its task inside the human body under the expertise of physicians and biologists, while the final outcome must be analyzed and coordinated by all the intervening research. A procedure starts after a specific need has been identified. Afterwards, an idea for a potential implant is created with the final insertion of the implant into a patient's body. The whole process appears to be very long because several stages have to be verified: material synthesis, design and manufacturing of the prosthesis, combined with multiple material tests, followed by biomedical evaluation. Finally, a potential biomaterial must also pass all necessary regulatory requirements¹.

The physical character of the majority of the available biomaterials is solid. Depending on their nature and composition, they are divided into four major groups: biometals, biopolymers, bioceramics and various blends thereof, called biocomposites. All of them play very important

roles in both replacement and regeneration of various human tissues; however, setting biometals, biopolymers and biocomposites aside, this paper is focused on bioceramics only. The bioceramic materials are designed to be in contact with living tissues and have experienced great development in the last 50 years. The medical needs of an increasingly aging population have driven a great deal of research work looking for new bioceramic materials to regenerate and repair living bones damaged by disease or trauma. For those specific clinical applications, mainly in orthopedics and dentistry, bioceramics are playing a key role.

The use of ceramic materials represents an evolution of many aspects of human history. Namely, many millennia ago, the possibility to store grains in ceramic receptacles allowed man to become a settler instead of a nomad hunter. Some centuries ago, the use of structural ceramics also brought great advances in the quality of life of people with the possibility of making clay bricks and tiles. Decades ago, ceramics generated a new revolution in the human way of life, with development of the functional ceramics as dielectrics, semiconductors, magnets, piezoelectrics, high-temperature superconductors and so on. In addition, ceramics plays an important role in improving the quality and length of human life through their use as biomaterials and in medical devices¹.

In general, ceramics are inorganic materials with a combination of ionic and covalent bonding. Therefore, they have high melting temperatures, low electricity and heat conduction, as well as relatively high hardness. Regarding their mechanical behavior, ceramic materials exhibit

* Corresponding author: sedorozhkin@yandex.ru

high compressive strengths, but very much lower tensile strengths. In addition, they are stiff materials, with a high Young's modulus, and brittle because failure takes place without plastic deformation. With regard to their surface properties, ceramics usually possess high wetting degrees and surface tensions, which favor adhesion of proteins, cells and other biological moieties. Furthermore, the ceramic surfaces can be treated to reach very high polish limits. Currently, many research efforts are devoted to ceramics with interconnected porosity.

Regarding their composition, the vast majority of inorganic compounds (metal haloids, metal oxides, metal chalcogenides, metal nitrides, metal phosphides, metal carbides, as well as various oxygen-containing salts of metals (sulfates, phosphates, nitrates, acetates, carbonates, silicates, etc.)) are classed as ceramics. However, the chemical elements used to manufacture bioceramics form just a small set in the Periodic Table. Namely, bioceramics might be prepared from alumina, zirconia, magnesia, carbon, silica-contained and calcium-contained compounds, as well as from a limited number of other chemicals. Therefore, all these compounds plus calcium phosphates, calcium sulfates, certain glasses and glass-ceramics appear to be genuine examples of bioceramics. Although carbon is not a compound but an element and conducts electricity in its graphite form, it is also considered as a ceramic owing to its many ceramic-like properties. Nowadays, new advanced bioceramics are under study, including ordered mesoporous silica materials or specific compositions of organic-inorganic hybrids. All these compounds might be manufactured in both dense and porous forms in bulk, as well as in the forms of crystals, powders, particles, granules, scaffolds and/or coatings¹.

II. General Knowledge and Definitions

Several definitions have been developed for the term "biomaterials". For example, by the end of the 20th century, the consensus developed by the experts was the following: biomaterials were defined as synthetic or natural materials to be used to replace parts of a living system or to function in intimate contact with living tissues². However, in September 2009, a more advanced definition was introduced: "A biomaterial is a substance that has been engineered to take a form which, alone or as part of a complex system, is used to direct, by control of interactions with components of living systems, the course of any therapeutic or diagnostic procedure, in human or veterinary medicine"³. Changes to the definition were accompanied by a shift in both the conceptual ideas and the expectations of biological performance, which mutually changed over time⁴.

In general, the biomaterials discipline is founded in the knowledge of the synergistic interaction of material science, biology, chemistry, medicine and mechanical science and it requires the input of comprehension from all these areas so that potential implants perform adequately in a living body and interrupt normal body functions as little as possible⁵. As biomaterials deal with all aspects of the material synthesis and processing, knowledge of chemistry, material science and engineering appear to be essential. On the other hand, since clinical implantology is the main pur-

pose of biomaterials, biomedical sciences become the key part of the research. These include cell and molecular biology, histology, anatomy and physiology. The final aim is to achieve the correct biological interaction of the artificial grafts with living tissues of a host. Thus, to achieve the goals, several stages must be performed, such as: material synthesis, design and manufacturing of prostheses, followed by various types of tests. Furthermore, before clinical applications, any potential biomaterial must also pass all regulatory requirements⁶.

In any case, biomaterials are intended to interface with biological systems *in vivo* to evaluate, treat, augment or replace any tissue, organ or function of the body and are now used in several different applications throughout the body. Thus, biomaterials are solely associated with the health care domain and must have an interface with tissues or tissue components. One should stress that any artificial materials that are simply in contact with skin, such as hearing aids and wearable artificial limbs, are not included in the definition of biomaterials since the skin acts as a protective barrier between the body and the external world^{7, 8}.

The major difference between biomaterials and other classes of materials lies in their ability to remain in a biological environment without damaging their surroundings nor being damaged in the process. Therefore, biomaterials must be distinguished from biological materials because the former are the materials that are accepted by living tissues and, therefore, they might be used for tissue replacements, while the latter are just the materials being produced by various biological systems (wood, cotton, bones, chitin, etc.)⁹. Furthermore, there are biomimetic materials, which are not made by living organisms but have the composition, structure and properties similar to those of biological materials. Concerning the subject of the current review, bioceramics (or biomedical ceramics) are defined as biomaterials having a ceramic origin. Now it is important to define the meaning of ceramics. According to Wikipedia, the free encyclopedia: "The word ceramic comes from the Greek word *κεραμικός* (*keramikos*), "of pottery" or "for pottery", from *κέραμος* (*keramos*), "potter's clay, tile, pottery". The earliest known mention of the root "ceram-" is the Mycenaean Greek *ke-ra-me-we*, "workers of ceramics", written in Linear B syllabic script. The word "ceramic" may be used as an adjective to describe a material, product or process, or it may be used as a noun, either singular, or, more commonly, as the plural noun "ceramics". A ceramic material is an inorganic, non-metallic, often crystalline oxide, nitride or carbide material. Some elements, such as carbon or silicon, may be considered as ceramics. Ceramic materials are brittle, hard, strong in compression, weak in shearing and tension. They withstand chemical erosion that occurs in other materials subjected to acidic or caustic environments. Ceramics can generally withstand very high temperatures, such as temperatures that range from 1000 °C to 1600 °C (1800 °F to 3000 °F). Glass is often not considered a ceramic because of its amorphous (non-crystalline) character. However, glassmaking involves several steps of the ceramic process and the mechanical properties of glass are similar to those of ceramic materials."¹⁰. Similar to any other type

of biomaterial, bioceramics can have structural functions as joint or tissue replacements, be used as coatings to improve biocompatibility, as well as function as resorbable lattices, providing temporary structures and frameworks that are dissolved and/or replaced as the body rebuilds the damaged tissues^{11–14}. Some types of bioceramics feature a drug-delivery capability^{15–18}.

most remarkable accomplishments of research, development, production and quality assurance by the end of the past century¹¹.

III. Brief Historical Overview

In medicine, bioceramics have been used for millennia to alleviate pain and restore functions of diseased or damaged calcified tissues (bones and teeth) of the body. For example, in 1972, Amadeo Bobbio discovered Mayan skulls, some of them more than 4000 years old, in which missing teeth had been replaced by nacre substitutes²¹. In addition, according to Wikipedia, literature dating back to 975 AD notes that calcium sulfate was useful for setting broken bones. However, those were ex vivo applications. According to the available literature, by the end of the 19th century, surgeons were already using plaster of Paris as a bone-filling substitute²². Nevertheless, it was a famous German surgeon Themistocles Gluck (1853–1942), who, amongst his range of contributions, on 20 May 1890 performed the first well-documented ivory (virtually pure biological apatite) knee replacement bedded in a calcium-sulfate-based cement, which was followed by a total wrist replacement in another patient three weeks later²³. Later in 1890, Gluck presented a further case of a total knee replacement to the Berlin Medical Society: at only 35 days after operation, the patient was free of pain with active knee flexion and extension. All the joint arthroplasties performed by Gluck were remarkably successful in the short term; however, all ultimately failed because of chronic infections^{24, 25}. With regard to other types of bioceramics, the first attempt to implant laboratory-produced calcium phosphate as an artificial material to repair surgically created defects in rabbit bones was performed in 1920²⁶ by the US surgeon Fred Houdlette Albee (1876–1945), who invented bone grafting²⁷ and some other advances in orthopedic surgery. Extensive studies of plaster of Paris to repair bone defects continued through the first half of 1900s^{28, 29}, while the first application of alumina as a biomaterial was suggested in 1933 by Rock³⁰.

As written in the literature, the “modern” era of bioceramics can be traced to Smith’s successful study of 1963 on a ceramic bone substitute material Cerosium®, composed of a porous aluminate ceramic impregnated with an epoxy resin. The porosity of that bioceramic was controlled at 48 % in analogy to a comparable value for natural bone and in order to produce net physical properties very close to those of bone³¹. In 1960s, several other publications on the application of the ceramic materials as prostheses were published as well^{32–35}. In 1969, the first scientific study of the outstanding biomedical properties of zirconia emerged³⁶. In 1971, bioactive glasses were prepared³⁷. In 1972, a famous paper by Boutin was published³⁸; since then alumina took off on its worldwide triumphal course as a suitable bioceramic for femoral balls of hip endoprostheses. Concerning the earliest appearance of term “bioceramics” in the scientific literature, according to the available databases, the first paper with this term in the abstract was published in 1971³⁹, while the earliest papers with the term in the title were published in 1972^{40, 41}. On 26 April 1988, the first international symposium on bioceramics

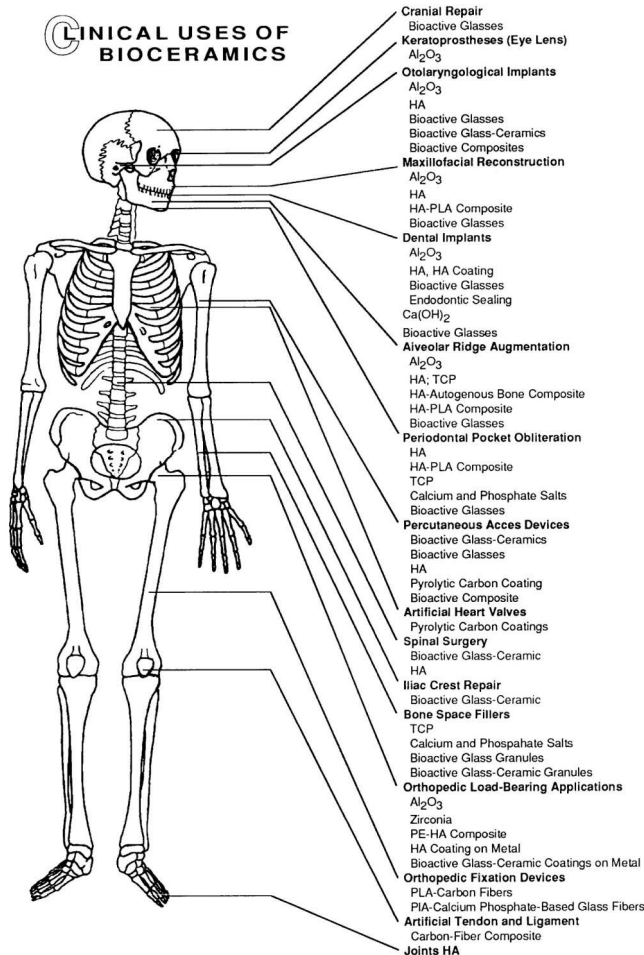


Fig. 1: Clinical uses of bioceramics. Reprinted from Ref. 19 with permission.

Bioceramics are produced in a variety of forms and phases and serve many different functions in the repair of the body. Fig. 1 presents a graphical sketch of clinical uses of bioceramics within the human body¹⁹. In biomedical applications, bioceramics are used in the form of bulk materials of a specific shape, called implants, prostheses or prosthetic devices. A great challenge facing its medical application is, first, to replace and, second, to regenerate old and deteriorating bones with a biomaterial that can be replaced by a new mature bone without transient loss of mechanical support^{8, 20}. Since the average lifespan of humans is now 80+ years and the major need for spare parts begins at about 60 years of age, the after-effects of the implanted bioceramics need to last, at least, for 20+ years. This demanding requirement of survivability is under conditions of use that are especially harsh to implanted biomaterials: corrosive saline solutions at 37 °C under variable, multiaxial and cyclical mechanical loads. The excellent performance of the specially designed bioceramics that have survived these clinical conditions represented one of the

was held in Kyoto, Japan. The historical development of bioceramics is schematically shown in Fig. 2⁴².

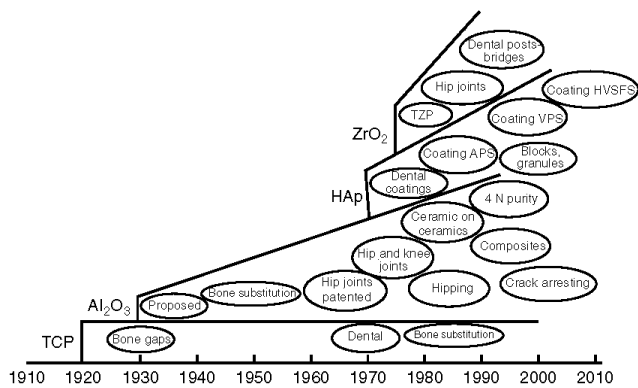


Fig. 2: Application of bioceramics in medical devices: 100 years of history. Reprinted from Ref. 42 with permission.

IV. The First, Second and Third Generations of Bioceramics

Both conceptually and historically, the study of biomaterials (therefore, bioceramics as well) can be divided into the first⁴³, second⁴⁴ and third⁴⁵ generations (Fig. 3)¹. The first generation of biomaterials started in the 1960s, when the goal was to minimize reactivity. Briefly, when synthetic materials were first used in biomedical applications, the requirements for use were suitable physical properties to match those of the replaced tissue with a minimal toxic response of the host, so biologically inert or nearly inert materials were used in order to reduce the corrosion and the releasing ions and particles after implantation to minimize the immune response and foreign body reaction. Mechanical properties and toxicity also played a leading role in the selection of materials for implant manufacture. Therefore, the first generation of biomaterials was used solely for tissue replacement. When inert biomaterials (strictly speaking, a material should never be considered as totally inert; such materials just do not create a direct interface with the adjacent tissues) are placed inside the body, they would elicit a foreign fibrous capsule around the material which isolates it from the surrounding tissue. This biological shielding leads to mechanical (stress) shielding, known to promote micro-motion and subsequent aseptic implant loosening. The representative examples of this type of bioceramics are alumina (Al₂O₃) and zirconia (ZrO₂). Owing to their high strength, excellent corrosion and wear resistances, stability, non-toxicity and in vivo biocompatibility, they are widely used to fabricate femoral heads⁴³. Non-oxide almost inert bioceramics such as silicon nitride (Si₃N₄)^{46,47} and silicon carbide (SiC)⁴⁸ are being developed as well.

In addition, there are certain compositions of ceramics that are able to form a mechanically strong bond to bones. These materials have become known as bioactive bioceramics. All of them represent the second generation of biomaterials, which uses the materials' ability to interact with the biological environment to enhance the biological response and provide the tissue/surface bonding. Among them, there is a group of bioresorbable biomaterials that possess an ability to degrade when tissues are re-

generated and healed. Thus, around the 1980s, the objective changed to obtain favorable interactions with the living body, namely a bioactive response or degradation⁴⁴. Therefore, the second generation of biomaterials is used for tissue regeneration. A common characteristic of this generation of biomaterials is a time-dependent, kinetic modification of the surface that occurs upon implantation as the result of interactions with the physiological fluids. Namely, in the case of bone grafts, a biologically active layer of carbonated apatite is formed on the surface of such biomaterials and this layer provides the bonding interface with adjacent bones and surrounding tissues. This carbonated apatite phase appears to be chemically and structurally equivalent to the mineral phase of bones which is responsible for the interfacial bonding¹¹. Moreover, owing to the actions of living cells, this apatite can form new bones. Specific compositions of calcium phosphates and/or sulfates, bioactive glasses and glass-ceramics are examples of the second-generation bioceramics used clinically for bone tissue augmentation. For biomedical applications, these materials are provided as powders, both porous and dense pieces, injectable mixtures, self-setting formulations and coatings. All of them have excellent features in terms of biocompatibility and bioactivity, but their mechanical properties are poor¹.

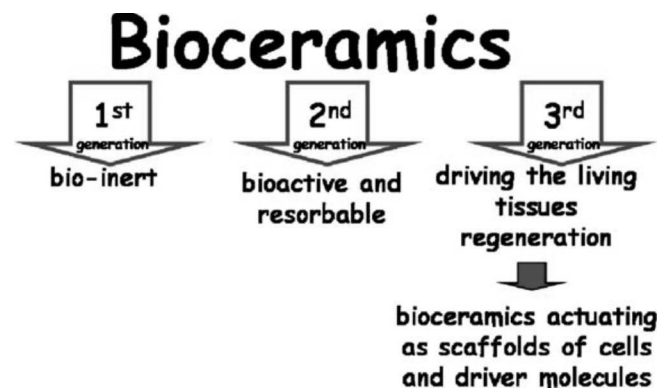


Fig. 3: A schematic layout of the three generations of bioceramics. Reprinted from Ref. 1 with permission.

The third generation of biomaterials uses both the bioactive and the bioresorbable materials as temporary three-dimensional porous structures (scaffolds) which are able to activate genes that stimulate regeneration of living tissue. For these biomaterials, concepts of bioactivity and biodegradability are combined, and this combination of both concepts appears to be the key feature for the third generation of biomaterials. Thus, the major purpose of the third generation of bioceramics is basically to provide an adequate scaffolding system which helps living cells to perform their natural processes⁴⁵. In addition, a concept of porosity and its range of order appears to be of paramount importance. Namely, bioceramics with mesoporosity between 2 and 50 nm are of interest for applications where drugs and/or biologically active molecules are loaded and later released to help in the bone regeneration process. Macroporous materials with pore dimensions exceeding several microns appear to be suitable as scaffolds for tissue engineering. The studies in third-generation bioceramics

are based more on biology and follow the purpose of tissue regeneration (rather than tissue replacement) with attempts to develop artificial materials able to restore damaged biological tissues in situations when the human body cannot perform this by itself. One attempt consists of designing biomimetic materials that combine synthetic materials with cellular recognizing positions. Generally, this category is based on the second-generation bioceramics with porosity, loaded with biologically active substances. The examples comprise mesoporous silica, mesoporous ordered glasses, porous calcium phosphate scaffolds and organic-inorganic hybrids with cellular recognizing positions¹.

To finalize this section, one should stress that the first generation of inert bioceramics was aimed at serving as artificial bone grafts, the second generation of bioactive and bioresorbable bioceramics was developed to mimic some biomineralization-related functions, while the purpose of the third generation of bioceramics is basically to provide an adequate scaffolding system that helps bone cells to perform their restorative functions. In addition, one should mention that the fourth generation of biomaterials was announced in 2016. According to the authors, it should be designed to both manipulate and monitor cellular bioelectrical signals⁴⁹. Currently, it has nothing in common with bioceramics.

V. Various Types of the Bioceramic/Issue Interfaces

In general, no implanted material appears to be totally inert for the surrounding tissues; thus, all implants elicit a response from the host tissues. This response occurs at the tissue/implant interface and depends on many factors. According to the literature¹⁹, all factors affecting this interfacial response may be divided into two groups: factors from the tissue side and those from the implant side. The first group of the factors comprises the type of tissue, the health of the tissue, the age of the tissue, blood circulation in both the tissue and at the interface, as well as motion at the interface, while the second group of the factors includes the implant composition, phases in the implant, phase boundaries, surface morphology, surface porosity and chemical reactions. Factors such as closeness of fit and a mechanical load appear to have an influence from the both sides of the tissue/implant interfaces¹⁹.

A combination of all the aforesaid factors creates the overall tissue response. According to the available knowledge on the subject, there are four possible types of tissue responses to the implanted materials: biotoxic, biologically nearly inert, bioactive and bioresorbable. A biotoxic response causes cell death in the surrounding tissues owing to release of dangerous chemicals that are able to migrate within tissue fluids and cause systemic damage to the patient. Since a lack of toxicity appears to be critical, biotoxic materials are excluded from any type of biomedical applications. Therefore, in the case of bioceramics, just three types of the implant/tissue responses – biologically nearly inert, bioactive and bioresorbable – are considered. It is important to note that these three types of responses correlate fully with the aforementioned three generations of bioceramics (Fig. 3). Thus, each generation of bioceramics appears to possess an implant/tissue response of its own.

VI. The Major Properties of Bioceramics

(1) Mechanical properties

The modern generation of bioceramics is designed to stimulate the body's own self-repairing abilities⁴⁵. Therefore, during healing, a mature bone should replace the modern grafts and this process must occur without transient loss of mechanical support. Unluckily for material scientists, a human body provides one of the most inhospitable environments for implanted biomaterials. It is warm, wet and both chemically and biologically active. For example, a diversity of body fluids in various tissues might have a solution pH varying from 1 to 9. In addition, a body is capable of generating quite massive force concentrations and the variance in such characteristics among individuals might be enormous. Typically, bones are subjected to ~ 4 MPa loads, whereas tendons and ligaments experience peak stresses in the range of 40–80 MPa. The hip joints are subjected to an average load up to three times body weight (3000 N) and peak loads experienced during jumping can be as high as ten times body weight. These stresses are repetitive and fluctuate depending on the nature of the activities, which can include standing, sitting, jogging, stretching and climbing. Therefore, all types of implants must sustain attacks of a great variety of aggressive conditions⁵⁰. Regrettably, there is presently no artificial material fulfilling all these requirements.

For dense bioceramics, the strength is a function of the grain sizes. Namely, finer grain size bioceramics have smaller flaws at the grain boundaries and thus are stronger than those with larger grain sizes. Thus, in general, the strength for ceramics is proportional to the inverse square root of the grain sizes⁵¹. In addition, the mechanical properties decrease significantly with increasing content of an amorphous phase, microporosity and grain sizes, while a high crystallinity, a low porosity and small grain sizes tend to give higher stiffness, higher compressive and tensile strength and greater fracture toughness. Furthermore, ceramic strength appears to be very sensitive to slow crack growth⁵². Accordingly, from the mechanical point of view, bioceramics appear to be brittle polycrystalline materials for which the mechanical properties are governed by crystallinity, grain size, grain boundaries, porosity and composition⁵³. Thus, they possess poor mechanical properties (for instance, low impact and fracture resistances) that do not allow bioceramics to be used in load-bearing areas, such as artificial teeth or bones^{11–14}. For example, fracture toughness (this is a property that describes the ability of a material containing a crack to resist fracture and is one of the most important properties of any material for virtually all design applications) of hydroxyapatite bioceramics does not exceed the value of ~ 1.2 MPa·m^{1/2}⁵⁴ (human bone: 2–12 MPa·m^{1/2}). It decreases exponentially with increasing porosity⁵⁵.

Furthermore, strength decreases almost exponentially with increasing porosity^{56, 57}. However, by changing the pore geometry, it is possible to influence the strength of porous bioceramics. It is also worth mentioning that porous bioceramics are considerably less fatigue-resistant

than dense bioceramics (in materials science, fatigue is the progressive and localized structural damage that occurs when a material is subjected to cyclic loading). Both grain sizes and porosity have been reported to influence the fracture path, which itself has little effect on the fracture toughness^{53,58}. However, no obvious decrease in mechanical properties was found after the bioceramics had been aged in the various solutions during the different periods of time⁵⁹.

Owing to a high brittleness (associated with low crack resistance), the biomedical applications of bioceramics are limited. Therefore, ways are continuously sought to improve their reliability. Namely, diverse reinforcements (ceramics, metals or polymers) have been applied to manufacture various biocomposites and hybrid biomaterials⁶⁰. Another approach to improve the mechanical properties of bioceramics is to cover the items with polymeric coatings^{61–63} or infiltrate porous structures with polymers^{64–66}.

(2) Electric/dielectric and piezoelectric properties

Occasionally, an interest in both electric/dielectric^{67–80} and piezoelectric^{81,82} properties of bioceramics is expressed. For example, a surface ionic conductivity of both porous and dense hydroxyapatite bioceramics was examined for humidity sensor applications, since the room temperature conductivity was influenced by relative humidity⁶⁸. Namely, the ionic conductivity of solid hydroxyapatite was a subject of research for its possible use as a gas sensor for alcohol⁶⁹, carbon dioxide^{67,76} or carbon monoxide⁷². Electric measurements were also used as a characterization tool to study the evolution of microstructure⁷⁰.

The electric properties of bioceramics appear to influence their biomedical applications. For example, there is interest in polarization of hydroxyapatite bioceramics to generate a surface charge by the application of electric fields at elevated temperatures^{83,84}. The presence of surface charges was shown to have a significant effect on both *in vitro* and *in vivo* crystallization of biological apatite^{85–91}. Furthermore, growth of both biomimetic calcium orthophosphates and bones was found to be accelerated on negatively charged surfaces and decelerated at positively charged surfaces^{89–102}. A similar effect was found for adsorption of bovine serum albumin¹⁰³. In addition, the electric polarization was found to accelerate a cytoskeleton reorganization of osteoblast-like cells^{104–107}, extend bioactivity¹⁰⁸, enhance bone ingrowth through the pores of porous implants¹⁰⁹ and influence cell activity^{110,111}.

(3) Possible transparency

Single crystals of many ceramic materials are optically transparent to visible light. Since bioceramics have a polycrystalline nature with a random orientation of large amounts of small crystals, it is opaque and white in color, unless colored dopants are added. However, in some cases, a transparency is convenient to provide some essential advantages (e.g. to enable direct viewing of living cells, their attachment, spreading, proliferation, and osteogenic differentiation cascade in a transmitted light). Thus, trans-

parent bioceramics (Fig. 4)¹¹² have been prepared and investigated^{112–121}. They can exhibit an optical transmittance of ~66% at a wavelength of 645 nm¹¹⁸. The preparation techniques include hot isostatic pressing¹²⁰, ambient-pressure sintering¹¹³, gel casting coupled with low-temperature sintering^{114,117}, pulse electric current sintering¹¹⁵, as well as spark plasma sintering^{122–128}. Fully dense, transparent bioceramics are obtained at temperatures above ~800 °C. Depending on the preparation technique, the transparent bioceramics have a uniform grain size and always are pore-free. Furthermore, translucent bioceramics are also known^{129–131}. Concerning possible biomedical applications, the ceramics that are optically transparent to visible light can be useful for direct viewing of other objects, such as cells, in some specific experiments¹¹⁶. In addition, bioceramics with transparency to laser light may appear to be convenient for minimal invasive surgery by allowing the laser beam to pass through it to treat the injured tissues located underneath. However, owing to a lack of both porosity and the great necessity to have see-through implants inside the body, the transparent and translucent forms of bioceramics will hardly be extensively used in medicine except the aforementioned cases and possible eye implants.



Fig. 4: Transparent hydroxyapatite bioceramics prepared by spark plasma sintering at 900 °C from nano-sized HA single crystals. Reprinted from Ref.¹¹² with permission.

(4) Porosity

Porosity is defined as a percentage of voids in solids and this morphological property is independent of the material. The surface area of porous bodies is much higher, which guarantees good mechanical fixation in addition to providing sites on the surface that allow chemical bonding between the bioceramics and bones¹³². Furthermore, a porous material may have both closed (isolated) pores and open (interconnected) pores. The latter look like tunnels and are accessible by gases, liquids and particulate suspensions¹³³. The open-cell nature of porous materials (also known as reticulated materials) is a unique characteristic essential in many applications. In addition, pore dimensions are also important. Namely, the dimensions of open pores are directly related to bone formation, since such pores grant both the surface and space for cell adhesion and bone ingrowth^{134–136}. On the other hand, pore interconnection provides the ways for cell distribution and migration, as well as allowing efficient *in vivo* blood vessel formation suitable for sustaining bone tissue neo-formation and possibly remodeling^{109,137–143}. Thus, porous bioceramics is colonized easily by cells and bone tissues^{137,143,144–151}. Therefore, interconnecting macroporosity (pore size > 100 μm)^{132,137,152,153} is intentionally introduced in solid bioceramics (Fig. 5). In

addition, macroporosity might be formed artificially due to a release of various easily removable compounds and, for that reason, incorporation of pore-creating additives (porogens) is the most popular technique to create macroporosity. The porogens are crystals, particles or fibers of either volatile (they evolve gases at elevated temperatures) or soluble substances. The popular examples comprise paraffin^{154–156}, naphthalene^{157–159}, sucrose^{160,161}, NaHCO₃^{162–164}, NaCl^{165,166}, polymethylmethacrylate^{167–169}, hydrogen peroxide^{170–175}. Several other compounds^{176–187} might be used as porogens as well. The ideal porogen should be nontoxic and be removed at ambient temperature, thereby allowing the bioceramic/porogen mixture to be injected directly into a defect site and allowing the scaffold to fit the defect¹⁸⁸. Sintering particles, preferably spheres of equal size, is a similar way to generate porous 3D bioceramics. However, pores resulting from this method are often irregular in size and shape and not fully interconnected with one another. Schematic drawings of various types of the ceramic porosity are shown in Fig. 6¹⁸⁹.

Many other techniques, such as replication of polymer foams by means of impregnation^{190–194} (Fig. 5), various types of casting^{175,195–203}, surfactant washing²⁰⁴,

microemulsions^{205,206}, ice templating^{207–210}, as well as many other approaches^{211–246} have been applied to fabricate porous bioceramics. In addition, both natural porous materials, such as coral skeletons^{247,248} or shells^{248,249}, and artificially prepared ones²⁵⁰ can be converted into porous bioceramics under the hydrothermal conditions (250 °C, 24–48 h) with the microstructure undamaged. Besides, porous bioceramics might be prepared by hardening of the self-setting formulations^{155,156,163,164,166,176,177,235}. In addition, porous bioceramics might be prepared by using different starting powders of the same compound and sintering at various temperatures by means of pressureless sintering²¹³. Porous bioceramics with improved strength might be fabricated from fibers or whiskers. In general, fibrous porous materials are known to exhibit improved strength owing to fiber interlocking, crack deflection and/or pull-out²⁵¹. Namely, porous bioceramics with well-controlled open pores were processed by sintering of fibrous particles²¹². Finally, superporous (~85 % porosity) bioceramics were developed, too^{231–233}. Additional information on the processing routes to produce porous ceramics can be found in the literature²⁵².

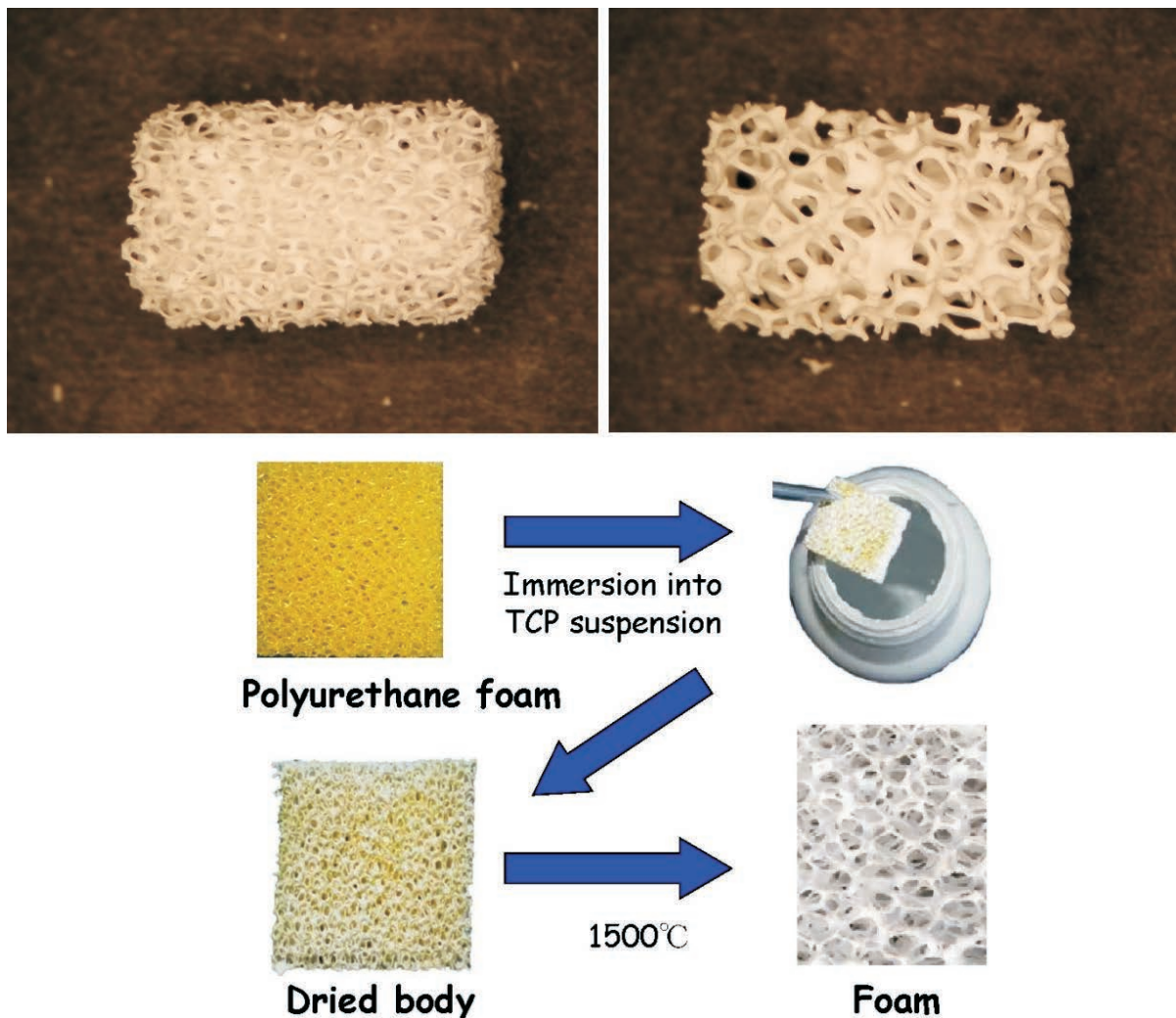


Fig. 5: Photographs of a commercially available porous bioceramics with different porosity (top) and a method of their production (bottom). For photos, the horizontal field width is 20 mm.

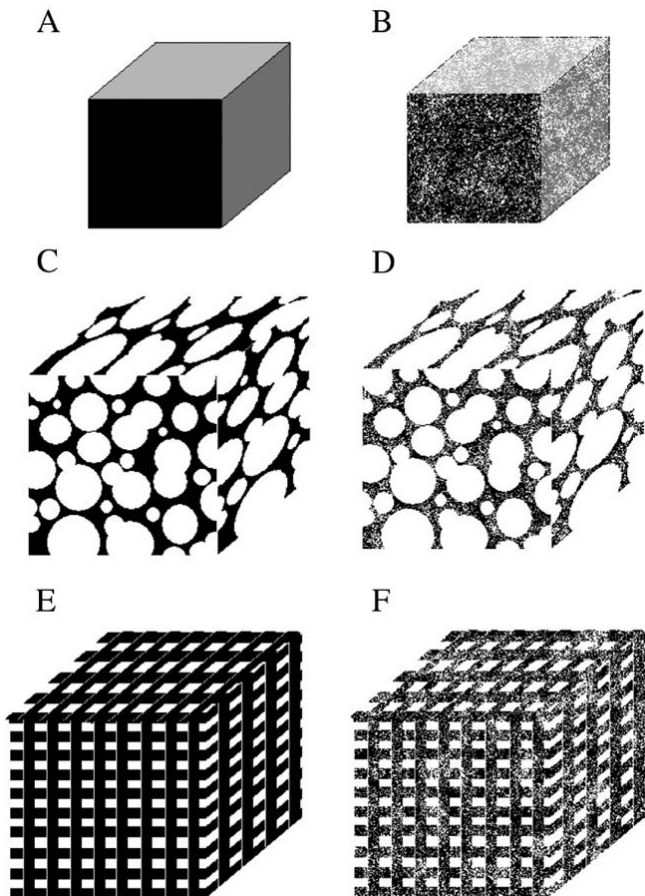


Fig. 6: Schematic drawings of various types of the ceramic porosity: A – non-porous, B – microporous, C – macroporous (spherical), D – macroporous (spherical) + micropores, E – macroporous (3D-printing), F – macroporous (3D-printing) + micropores. Reprinted from Ref. 189 with permission.

Bioceramic microporosity (pore size $< 10 \mu\text{m}$), which is defined by its capacity to be impregnated by biological fluids²⁵³, results from the sintering process, while the pore dimensions mainly depend on the material composition, thermal cycle and sintering time. The microporosity provides both a greater surface area for protein adsorption and increased ionic solubility. For example, embedded osteocytes distributed throughout microporous rods might

form a mechanosensory network, which would not be possible in scaffolds without microporosity^{254,255}. Bioceramics with nano-dimensional ($< 100 \text{ nm}$) pores might be fabricated as well^{256–260}. It is important to stress that differences in porogens usually influence the bioceramics' macroporosity, while differences in sintering temperatures and conditions affect the percentage of microporosity. Usually, the higher the sintering temperature is, the lower both the microporosity content and the specific surface area of bioceramics are. Namely, hydroxyapatite bioceramics sintered at $\sim 1200 \text{ }^\circ\text{C}$ shows significantly less microporosity and a dramatic change in crystal sizes if compared with that sintered at $\sim 1050 \text{ }^\circ\text{C}$ (Fig. 7)²⁶¹. Furthermore, the average shape of pores was found to transform from strongly oblate to round at higher sintering temperatures²⁶². The total porosity (macroporosity + microporosity) of bioceramics was reported to be $\sim 70 \%$ ²⁶³ or even $\sim 85 \%$ ^{231–233} of the entire volume. In the case of coralline hydroxyapatite or bovine-derived apatites, the porosity of the original biologic material (coral or bovine bone) is usually preserved during processing²⁶⁴. To finalize the production topic, creation of the desired porosity in bioceramics is a rather complicated engineering task and the interested readers are referred to the additional publications on the subject^{57, 136, 234, 265–273}.

Regarding the biomedical importance of porosity, studies revealed that increase of both the specific surface area and pore volume of bioceramics might greatly accelerate the *in vivo* process of apatite deposition and, therefore, enhance the bone-forming bioactivity. More importantly, precise control over the porosity, pore dimensions and internal pore architecture of bioceramics on different length scales is essential for understanding the structure-bioactivity relationship and the rational design of better bone-forming biomaterials^{271, 274, 275}. Namely, in antibiotic charging experiments, bioceramics with nano-dimensional ($< 100 \text{ nm}$) pores showed a much higher charging capacity ($1621 \mu\text{g/g}$) than that of a commercially available bioceramic ($100 \mu\text{g/g}$), which did not contain nano-dimensional porosity²⁶⁷. In other experiments, porous blocks were found to be viable carriers with sustained release profiles for drugs²⁷⁶ and antibiotics over

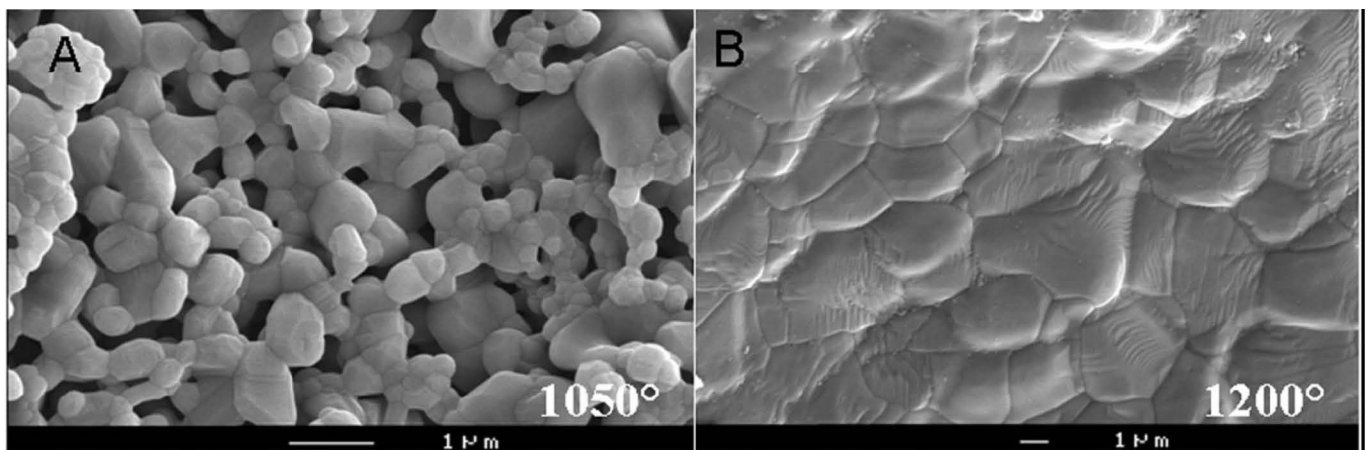


Fig. 7: SEM pictures of hydroxyapatite bioceramics sintered at (A) $1050 \text{ }^\circ\text{C}$ and (B) $1200 \text{ }^\circ\text{C}$. Note the presence of microporosity in A and not in B. Reprinted from Ref. 261 with permission.

12 days²⁷⁷ and 12 weeks²⁷⁸, respectively. Unfortunately, porosity significantly reduces the strength of implants⁵⁸. Thus, porous implants cannot be loaded and are used to fill only small bone defects. However, their strength increases gradually when bones ingrow into the porous network of implants^{279–282}. For example, bending strengths of 40–60 MPa for porous implants filled with 50–60% cortical bone were reported²⁷⁹, while in another study ingrown bone tissues increased strength of porous bioceramics by a factor of 3 to 4²⁸¹.

(5) Bioceramic scaffolds

Philosophically, the increase in life expectancy requires biological solutions to all biomedical problems, including orthopedic ones, which were previously managed with mechanical solutions. Therefore, since the end of 1990s, biomaterials research focuses on tissue regeneration instead of tissue replacement²⁸³. The alternatives include use of hierarchical bioactive scaffolds to engineer in vitro living cellular constructs for transplantation or use of bioresorbable bioactive particulates or porous networks to activate in vivo the mechanisms of tissue regeneration^{284,285}. Thus, the aim of bioceramics is to prepare artificial porous scaffolds able to provide the physical and chemical cues to guide cell seeding, differentiation and assembly into 3D tissues of a newly formed bone. Particle sizes, shape and surface roughness of the scaffolds are known to affect cellular adhesion, proliferation and phenotype^{286–291}. Additionally, the surface energy might play a role in attracting particular proteins to the bioceramic surface and, in turn, this will affect the cells' affinity to the material. More to the point, cells are exceedingly sensitive to the chemical composition and their bone-forming functions can be dependent on grain morphology of the scaffolds. For example, osteoblast functions were found to increase on nanodimensional fibers compared to nanodimensional spheres because the former more closely approximated the shape of biological apatite in bones²⁹². Besides, a significantly higher osteoblast proliferation on hydroxyapatite bioceramics sintered at 1200 °C as compared to bioceramics sintered at 800 °C and 1000 °C was reported²⁹³. A schematic drawing of the key scaffold properties affecting a cascade of biological processes occurring after implantation of calcium phosphate bioceramics is shown in Fig. 8²⁹⁴.

VII. Clinical Experience

To date, not many publications are known on clinical application of cell-seeded bioceramic scaffolds for bone tissue engineering in humans. Namely, Quarto et al.²⁹⁵ were the first to report a treatment of large (4–7 cm) bone defects of the tibia, ulna and humerus in three patients from 16 to 41 years old, where the conventional surgical therapies had failed. The authors implanted a custom-made unresorbable porous hydroxyapatite bioceramics seeded with in vitro expanded autologous bone marrow stromal cells. In all three patients, radiographs and computed tomographic scans revealed abundant callus formation along the implants and good integration at the interfaces with the host bones by the second month after surgery²⁹⁵. In the same year, Vacanti et al.²⁹⁶ reported the case of a man

who had a traumatic avulsion of the distal phalanx of a thumb. The phalanx was replaced with a specially treated natural coral (porous hydroxyapatite; 500-pore ProOsteon) implant that was previously seeded with in vitro expanded autologous periosteal cells. The procedure resulted in the functional restoration of a stable and biomechanically sound thumb of normal length, without the pain and complications that are usually associated with harvesting a bone graft.

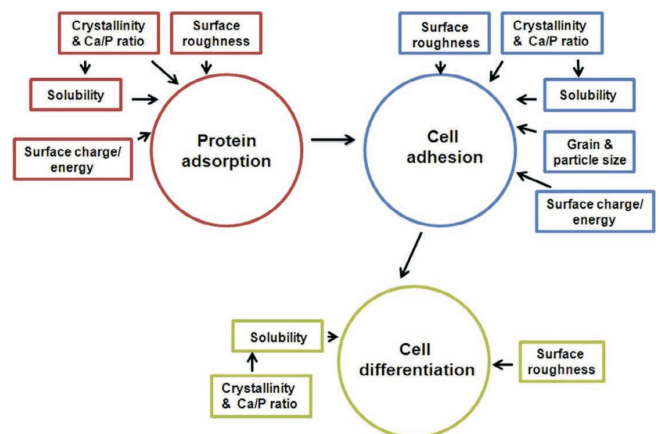


Fig. 8: A schematic drawing of the key scaffold properties affecting a cascade of biological processes occurring after implantation of calcium phosphate bioceramics. Reprinted from Ref.²⁹⁴ with permission.

Morishita et al.²⁹⁷ treated a defect resulting from surgery of benign bone tumors in three patients using porous hydroxyapatite bioceramics seeded with in vitro expanded autologous bone marrow stromal cells after osteogenic differentiation of the cells. Two bone defects in a tibia and one defect in a femur were treated. Although ectopic implants in nude mice were mentioned to show the osteogenicity of the cells, details such as the percentage of the implants containing bone and at what quantities were not reported. Furthermore, cell-seeded scaffolds were found to be superior to autograft, allograft or cell-seeded allograft in terms of bone formation at ectopic implantation sites²⁹⁸.

An innovative appliance named the stem cell screen-enrich-combine(-biomaterials) circulating system (SECCS) was designed in another study²⁹⁹. In that study, 42 patients who required bone graft underwent SECCS-based treatment. Their bone marrow samples and calcium phosphate granules were processed in the SECCS for 10–15 minutes, to produce composites. These composites were grafted back into bone defect sites. The results showed 85.53% ± 7.95% autologous mesenchymal stem cells were successfully screened, enriched, and seeded on the bioceramic scaffolds synchronously. Clinically, all patients obtained satisfactory bone healing²⁹⁹.

Besides, it has been hypothesized that dental follicle cells combined with calcium phosphate scaffolds might become a novel therapeutic strategy to restore periodontal defects³⁰⁰. In yet another study, the behavior of human periodontal ligament stem cells on a hydroxyapatite-coated genipin-chitosan scaffold in vitro was studied followed

by evaluation on bone repair in vivo³⁰¹. The study demonstrated the potential of this formulation for bone regeneration.

To conclude this section, one must mention that bio-ceramic scaffolds are also used in veterinary orthopedics to promote animal bone healing in areas in which bony defects exist^{302,303}.

VIII. Conclusions

Bioceramics have already become an integral and vital segment of our modern health care system. Therefore, in this section, the general information on the subject has been collected and summarized. Briefly, among the available types of bioactive and bioresorbable bioceramics, both pure and ion-substituted calcium phosphates and related composites currently are largely used for bone regeneration applications. These materials offer a large variety of compositions and/or structures according to their stoichiometry, the substitution elements' nature and content, the crystallinity and the crystal dimensions with the variable and adjustable osteoconductive and/or osteoinductive properties. At present, calcium phosphates are commercially available in different dimensions and shapes such as powders, granules, porous scaffolds, coatings, injectable and self-setting formulations. In addition, for a long time after their discovery by Hench in 1969, various types of bioactive glasses have also been commercialized for bone grafting purposes, while the recent developments are mainly devoted to sol-gel processing allowing achievement of larger composition ranges at lower temperature treatment and to the performance of coatings on various substrates. The subsequent development of bioactive glass ceramics was carried out to enhance the mechanical properties of bioglasses and led to satisfactory commercial products for small-bone replacements²⁸².

Regarding bioinert bioceramics, they are currently used as permanent load-bearing parts and comprise inert oxides like alumina, stabilized zirconia, spinel, related micro or nano-composites and, since recently, non-oxide bioceramics such as silicon nitride and carbide. Many different products prepared from alumina, zirconia and composites are successfully applied for dental restoration and orthopedic devices. The main innovations in progress concern two different aspects: the material and the processing techniques. Concerning the material aspect, research is focused on the development of a particular microstructure favorable for better mechanical properties, the use of new stabilizer ions for tetragonal zirconia and cermet compositions. For the fabrication methods, the novelties concern both additive and subtractive manufacturing techniques recently applied to ceramics materials and a promising reduction in time consumption²⁸².

Nevertheless, in all known fields of bioceramics, in order to induce higher bioreactivity, improved mechanical properties and/or better localized drug delivery abilities, a general tendency to used nano-scaled particles and/or grains can be observed. Thus, elaboration and manufacturing of the nano-dimensional and nano-crystalline bioceramics appear to be the hot point of current research and development.

Compliance with Ethical Standards

There are no potential conflicts of interest.

There is no research involving human participants and/or animals (this is a review).

There is no informed consent because I am a single author.

References

- Vallet-Regí, M.: Evolution of bioceramics within the field of biomaterials, *C. R. Chim.*, **13**, 174–185, (2010).
- Williams, D.F.: The Williams dictionary of biomaterials. Liverpool University Press, Liverpool, UK, 368, (1999).
- Williams, D.F.: On the nature of biomaterials. *Biomaterials*, **30**, 5897–5909, (2009).
- Bongio, M., van den Beucken, J.J.J.P., Leeuwenburgh, S.C.G., Jansen, J.A.: Development of bone substitute materials: From 'biocompatible' to 'instructive'. *J. Mater. Chem.*, **20**, 8747–8759, (2010).
- Mann, S. (Ed.): Biomimetic materials chemistry. Wiley-VCH, UK, 400, (1996).
- Vallet-Regí, M.: Bioceramics: Where do we come from and which are the future expectations, *Key Eng. Mater.*, **377**, 1–18, (2008).
- Ratner, B.D., Hoffman, A.S., Schoen, F.J., Lemons, J.E. (Eds.): Biomaterials science: An introduction to materials in medicine. 3rd edition. Academic Press, Oxford, UK, 1573, (2013).
- Ducheyne, P., Healy, K., Huttmacher, D.E., Grainger, D.W., Kirkpatrick, C.J. (Eds.): Comprehensive Biomaterials II. 2nd Edition. Seven-volume set. Elsevier, Amsterdam, Netherlands, 4858, (2017).
- Meyers, M.A., Chen, P.Y., Lin, A.Y.M., Seki, Y.: Biological materials: Structure and mechanical properties. *Prog. Mater. Sci.* **53**, 1–206, (2008).
- <https://en.wikipedia.org/wiki/Ceramic> (accessed in March 2018).
- Hench, L.L.: Bioceramics, *J. Am. Ceram. Soc.* **81**, 1705–1728, (1998).
- Hench, L.L., Day, D.E., Höland, W., Rheinberger, V.M.: Glass and medicine, *Int. J. Appl. Glass Sci.*, **1**, 104–117, (2010).
- Pinchuk, N.D., Ivanchenko, L.A.: Making calcium phosphate biomaterials. *Powder Metall. Metal Ceram.*, **42**, 357–371, (2003).
- Heimann, R.B.: Materials science of crystalline bioceramics: a review of basic properties and applications, *CMU J.* **1**, 23–46, (2002).
- Tomoda, K., Ariizumi, H., Nakaji, T., Makino, K.: Hydroxypatite particles as drug carriers for proteins, *Colloid Surf. B*, **76**, 226–235, (2010).
- Zamoume, O., Thibault, S., Regnié, G., Mecherri, M.O., Fiallo, M., Sharrock, P.: Macroporous calcium phosphate ceramic implants for sustained drug delivery, *Mater. Sci. Eng. C*, **31**, 1352–1356, (2011).
- Bose, S., Tarafder, S.: Calcium phosphate ceramic systems in growth factor and drug delivery for bone tissue engineering: a review, *Acta Biomater.*, **8**, 1401–1421, (2012).
- Arcos, D., Vallet-Regí, M.: Bioceramics for drug delivery, *Acta Mater.*, **61**, 890–911, (2013).
- Hench, L.L. (Ed.): An introduction to bioceramics. 2nd Ed. Imperial College Press, London, UK, 620, (2013).
- Ratner, B.D., Hoffman, A.S., Schoen, F.J., Lemons, J.E. (Eds.): Biomaterials science: An introduction to materials in medicine. 3rd edition. Academic Press, Oxford, UK, 1573, (2013).
- Bobbio, A.: The first endosseous alloplastic implant in the history of man, *Bull. Hist. Dent.*, **20**, 1–6, (1972).

- 22 Dreesmann, H.: About bone sealing, in german, *Beitr. Klin. Chir.*, **9**, 804–810, (1892).
- 23 Gluck, T.: Presentation on modern surgery, in german, *Langenbecks Arch. Klin. Chir.* **41**, 187–239, (1891).
- 24 Muster, D.: Themistocles gluck, berlin 1890: a pioneer of multidisciplinary applied research into biomaterials for endoprostheses, *Bull. Hist. Dent.*, **38**, 3–6, (1990).
- 25 Eynon-Lewis, N.J., Ferry, D., Pearse, M.F.: Themistocles Gluck: an unrecognised genius, *BMJ*, **305**, 1534–1536, (1992).
- 26 Albee, F.H., assisted by Morrison, H.F.: Studies in bone growth triple calcium phosphate as a stimulus to osteogenesis, *Ann Surg.*, **71**, 32–39, (1920).
- 27 Albee, F.H.: Bone-graft surgery. W.B. Saunders Company, Philadelphia and London, 417, (1915).
- 28 Edberg, E.: Some experiences of filling osseous cavities with plaster, *Acta Chir. Scand.* **67**, 313–319, (1930).
- 29 Peltier, L.F., Bickel, E.Y., Lillo, R., Thein, M.S.: The use of plaster of paris to fill defects in bone, *Ann. Surg.*, **146**, 61–69, (1957).
- 30 Rock, M.: Artificial spare parts for the interior and exterior of the human and animal body, in German, *Deutsches Reichspatent DRP 583 589*, 24 August 1933,
- 31 Smith, L.: Ceramic-plastic material as a bone substitute, *Arch. Surg.* **87**, 653–661 (1963).
- 32 Sandhaus, S.: Bone implants, and drills and tapes for bone surgery. EP 1083769 (A), application date Mar. 11, 1966, UK.
- 33 Rivault, M.A.: Evolution, conception and technology of fixed prosthesis made of ceramic and metal, in french, *Rev. Fr. Odontostomatol.*, **13**, 1367–1402, (1966).
- 34 Dumont, A., Appel, M., Favard, E.: Multiple prostheses made of ceramics on metal. soldering and artifacts of the junction, *Ann. Odontostomatol. (Lyon)*, **25**, 231–240, (1968).
- 35 Hulbert, S.F., Young, F.A., Mathews, R.S., Klawitter, J.J., Talbert, C.D., Stelling, F.H.: Potential of ceramic materials as permanently implantable skeletal prostheses, *J. Biomed. Mater. Res.*, **4**, 433–456, (1970).
- 36 Helmer, J.D., Driskell, T.D.: Research on bioceramics. Symposium on use of ceramics as surgical implants. Clemson University, Clemson, SC, USA 1969,
- 37 Hench, L.L., Splinter, R.J., Allen, W.C., Greenlee, T.K.: Bonding mechanisms at the interface of ceramic prosthetic materials, *J. Biomed. Mater. Res.*, **5**, 117–141, (1971).
- 38 Boutin, P.: Total hip arthroplasty with sintered alumina prosthesis, in french, *Rev. Chir. Orthop.*, **58**, 229–246, (1972).
- 39 Blakeslee, K.C., Condrate, R.A., Sr.: Vibrational spectra of hydrothermally prepared hydroxyapatites, *J. Am. Ceram. Soc.*, **54**, 559–563, (1971).
- 40 Garrington, G.E., Lightbody, P.M.: Bioceramics and dentistry. *J. Biomed. Mater. Res.* **6**, 333–343, (1972).
- 41 Cini, L., Sandrolini, S., Paltrinieri, M., Pizzoferrato, A., Trentani, C.: Bioceramic materials for replacement purposes. preliminary note, in italian, *Chir. Organi. Mov.*, **60**, 423–430, (1972).
- 42 Heimann, R.B., Lehmann, H.D.: Bioceramic coatings for medical implants: Trends and techniques. Wiley-VCH Verlag GmbH, Weinheim, Germany, 496, (2015).
- 43 Hench, L.L.: *Biomaterials, Science*, **208**, 826–831, (1980).
- 44 Hench, L.L., Wilson, J.: Surface-active biomaterials, *Science*, **226**, 630–636, (1984).
- 45 Hench, L.L., Polak, J.M.: Third-generation biomedical materials, *Science*, **295**, 1014–1017, (2002).
- 46 Bal, B.S., Rahaman, M.N.: Orthopedic applications of silicon nitride ceramics, *Acta Biomater.*, **8**, 2889–2898, (2012).
- 47 Pezzotti, G., McEntire, B.J., Bock, R.M., Boffelli, M., Zhu, W.L., Vitale, E., Puppulin, L., Adachi, T., Yamamoto, T., Kanamura, N., Bal, B.S.: Silicon nitride: a synthetic mineral for vertebrate biology, *Sci. Rep.*, **6**, 31717, (2016).
- 48 Rade, K., Martinčič, A., Novak, S., Kobe, S.: Feasibility study of SiC-ceramics as a potential material for bone implants, *J. Mater. Sci.*, **48**, 5295–5301, (2013).
- 49 Ning, C., Zhou, L., Tan, G.: Fourth-generation biomedical materials, *Mater. Today*, **19**, 2–3, (2016).
- 50 Black, J.: *Biological performance of materials: Fundamentals of biocompatibility*. Fourth Ed. CRC Press, Boca Raton, FL, USA, 520, (2005).
- 51 Carter, C.B., Norton, M.G.: *Ceramic materials: Science and engineering*. 2nd ed. Springer, New York, USA, 766, (2013).
- 52 Benaqqa, C., Chevalier, J., Saâdaoui, M., Fantozzi, G.: Slow crack growth behaviour of hydroxyapatite ceramics, *Biomaterials*, **26**, 6106–6112, (2005).
- 53 Pecqueux, F., Tancret, F., Payraudeau, N., Bouler, J.M.: Influence of microporosity and macroporosity on the mechanical properties of biphasic calcium phosphate bioceramics: modelling and experiment, *J. Eur. Ceram. Soc.*, **30**, 819–829, (2010).
- 54 Ramesh, S., Tan, C.Y., Sopyan, I., Hamdi, M., Teng, W.D.: Consolidation of nanocrystalline hydroxyapatite powder, *Sci. Technol. Adv. Mater.* **8**, 124–130, (2007).
- 55 Wagoner Johnson, A.J., Herschler, B.A.: A review of the mechanical behavior of CaP and CaP/polymer composites for applications in bone replacement and repair, *Acta Biomater.* **7**, 16–30, (2011).
- 56 Bouler, J.M., Trecant, M., Delecrin, J., Royer, J., Passuti, N., Daculsi, G.: Macroporous biphasic calcium phosphate ceramics: influence of five synthesis parameters on compressive strength, *J. Biomed. Mater. Res.*, **32**, 603–609, (1996).
- 57 Tancret, F., Bouler, J.M., Chamousset, J., Minois, L.M.: Modelling the mechanical properties of microporous and macroporous biphasic calcium phosphate bioceramics, *J. Eur. Ceram. Soc.*, **26**, 3647–3656, (2006).
- 58 le Huec, J.C., Schaefferbeke, T., Clement, D., Faber, J., le Rebellier, A.: Influence of porosity on the mechanical resistance of hydroxyapatite ceramics under compressive stress, *Biomaterials*, **16**, 113–118, (1995).
- 59 Hsu, Y.H., Turner, I.G., Miles, A.W.: Mechanical properties of three different compositions of calcium phosphate bioceramic following immersion in Ringer's solution and distilled water, *J. Mater. Sci. Mater. Med.*, **20**, 2367–2374, (2009).
- 60 Dorozhkin, S.V.: Calcium orthophosphate-containing biocomposites and hybrid biomaterials for biomedical applications, *J. Funct. Biomater.*, **6**, 708–832, (2015).
- 61 Dorozhkin, S.V., Ajaal, T.: Toughening of porous bioceramic scaffolds by bioresorbable polymeric coatings, *Proc. Inst. Mech. Eng. H.*, **223**, 459–470, (2009).
- 62 Dorozhkin, S.V., Ajaal, T.: Strengthening of dense bioceramic samples using bioresorbable polymers – a statistical approach, *J. Biomim. Biomater. Tissue Eng.*, **4**, 27–39, (2009).
- 63 Dressler, M., Dombrowski, F., Simon, U., Börnstein, J., Hodoroaba, V.D., Feigl, M., Grunow, S., Gildenhaar, R., Neumann, M.: Influence of gelatin coatings on compressive strength of porous hydroxyapatite ceramics, *J. Eur. Ceram. Soc.*, **31**, 523–529, (2011).
- 64 Martinez-Vazquez, F.J., Perera, F.H., Miranda, P., Pajares, A., Guiberteau, F.: Improving the compressive strength of bioceramic robocast scaffolds by polymer infiltration, *Acta Biomater.*, **6**, 4361–4368, (2010).
- 65 Fedotov, A.Y., Bakunova, N.V., Komlev, V.S., Barinov, S.M.: High-porous calcium phosphate bioceramics reinforced by chitosan infiltration, *Dokl. Chem.*, **439**, 233–236, (2011).
- 66 Martínez-Vázquez, F.J., Pajares, A., Guiberteau, F., Miranda, P.: Effect of polymer infiltration on the flexural behav-

- ior of β -tricalcium phosphate robocast scaffolds, *Materials*, **7**, 4001–4018, (2014).
- 67 Yamashita, K., Owada, H., Umegaki, T., Kanazawa, T., Futagamu, T.: Ionic conduction in apatite solid solutions, *Solid State Ionics*, **28–30**, 660–663, (1988).
- 68 Nagai, M., Nishino, T.: Surface conduction of porous hydroxyapatite ceramics at elevated temperatures, *Solid State Ionics*, **28–30**, 1456–1461, (1988).
- 69 Valdes, J.J.P., Rodriguez, A.V., Carrio, J.G.: Dielectric properties and structure of hydroxyapatite ceramics sintered by different conditions. *J. Mater. Res.* **10**, 2174–2177, (1995).
- 70 Fanovich, M.A., Castro, M.S., Lopez, J.M.P.: Analysis of the microstructural evolution in hydroxyapatite ceramics by electrical characterization, *Ceram. Int.*, **25**, 517–522, (1999).
- 71 Bensaoud, A., Bouhaouss, A., Ferhat, M.: Electrical properties in compressed poorly crystalline apatite, *J. Solid State Electrochem.*, **5**, 362–365, (2001).
- 72 Mahabole, M.P., Aiyer, R.C., Ramakrishna, C.V., Sreedhar, B., Khairnar, R.S.: Synthesis, characterization and gas sensing property of hydroxyapatite ceramic, *Bull. Mater. Sci.*, **28**, 535–545, (2005).
- 73 Tanaka, Y., Takata, S., Shimoe, K., Nakamura, M., Nagai, A., Toyama, T., Yamashita, K.: Conduction properties of non-stoichiometric hydroxyapatite whiskers for biomedical use, *J. Ceram. Soc. Jpn.*, **116**, 815–821, (2008).
- 74 Tanaka, Y., Nakamura, M., Nagai, A., Toyama, T., Yamashita, K.: Ionic conduction mechanism in Ca-deficient hydroxyapatite whiskers, *Mater. Sci. Eng. B*, **161**, 115–119, (2009).
- 75 Wang, W., Itoh, S., Yamamoto, N., Okawa, A., Nagai, A., Yamashita, K.: Electrical polarization of β -tricalcium phosphate ceramics, *J. Am. Ceram. Soc.*, **93**, 2175–2177, (2010).
- 76 Mahabole, M.P., Mene, R.U., Khairnar, R.S.: Gas sensing and dielectric studies on cobalt doped hydroxyapatite thick films, *Adv. Mater. Lett.*, **4**, 46–52, (2013).
- 77 Horiuchi, N., Nakaguki, S., Wada, N., Nozaki, K., Nakamura, M., Nagai, A., Katayama, K., Yamashita, K.: Polarization-induced surface charges in hydroxyapatite ceramics, *J. Appl. Phys.*, **116**, 014902, (2014).
- 78 Tofail, S.A.M., Gandhi, A.A., Gregor, M., Bauer, J.: Electrical properties of hydroxyapatite, *Pure Appl. Chem.*, **87**, 221–229, (2015).
- 79 Kaygili, O., Keser, S., Ates, T., Kirbag, S., Yakuphanoglu, F.: Dielectric properties of calcium phosphate ceramics, *Medziagotyra*, **22**, 65–69, (2016).
- 80 Suresh, M.B., Biswas, P., Mahender, V., Johnson, R.: Comparative evaluation of electrical conductivity of hydroxyapatite ceramics densified through ramp and hold, spark plasma and post sinter hot isostatic pressing routes, *Mater. Sci. Eng. C* **70**, 364–370, (2017).
- 81 Gandhi, A.A., Wojtas, M., Lang, S.B., Kholkin, A.L., Tofail, S.A.M.: Piezoelectricity in poled hydroxyapatite ceramics, *J. Am. Ceram. Soc.*, **97**, 2867–2872, (2014).
- 82 Bystrov, V.S.: Piezoelectricity in the ordered monoclinic hydroxyapatite, *Ferroelectrics*, **475**, 148–153, (2015).
- 83 Nakamura, S., Takeda, H., Yamashita, K.: Proton transport polarization and depolarization of hydroxyapatite ceramics, *J. Appl. Phys.*, **89**, 5386–5392, (2001).
- 84 Gittings, J.P., Bowen, C.R., Turner, I.G., Baxter, F.R., Chaudhuri, J.B.: Polarisation behaviour of calcium phosphate based ceramics, *Mater. Sci. Forum*, **587–588**, 91–95, (2008).
- 85 Itoh, S., Nakamura, S., Kobayashi, T., Shinomiya, K., Yamashita, K., Itoh, S.: Effect of electrical polarization of hydroxyapatite ceramics on new bone formation, *Calcif. Tissue Int.*, **78**, 133–142, (2006).
- 86 Iwasaki, T., Tanaka, Y., Nakamura, M., Nagai, A., Hashimoto, K., Toda, Y., Katayama, K., Yamashita, K.: Rate of bonelike apatite formation accelerated on polarized porous hydroxyapatite, *J. Am. Ceram. Soc.*, **91**, 3943–3949, (2008).
- 87 Itoh, S., Nakamura, S., Kobayashi, T., Shinomiya, K., Yamashita, K.: Enhanced bone ingrowth into hydroxyapatite with interconnected pores by electrical polarization, *Biomaterials*, **27**, 5572–5579, (2006).
- 88 Kobayashi, T., Itoh, S., Nakamura, S., Nakamura, M., Shinomiya, K., Yamashita, K.: Enhanced bone bonding of hydroxyapatite-coated titanium implants by electrical polarization, *J. Biomed. Mater. Res. A*, **82A**, 145–151, (2007).
- 89 Bodhak, S., Bose, S., Bandyopadhyay, A.: Role of surface charge and wettability on early stage mineralization and bone cell-materials interactions of polarized hydroxyapatite, *Acta Biomater.*, **5**, 2178–2188, (2009).
- 90 Sagawa, H., Itoh, S., Wang, W., Yamashita, K.: Enhanced bone bonding of the hydroxyapatite/ β -tricalcium phosphate composite by electrical polarization in rabbit long bone, *Artif. Organs*, **34**, 491–497, (2010).
- 91 Ohba, S., Wang, W., Itoh, S., Nagai, A., Yamashita, K.: Enhanced effects of new bone formation by an electrically polarized hydroxyapatite microgranule/platelet-rich plasma composite gel, *Key Eng. Mater.*, **529–530**, 82–87, (2013).
- 92 Yamashita, K., Oikawa, N., Umegaki, T.: Acceleration and deceleration of bone-like crystal growth on ceramic hydroxyapatite by electric poling, *Chem. Mater.*, **8**, 2697–2700, (1996).
- 93 Teng, N.C., Nakamura, S., Takagi, Y., Yamashita, Y., Ohgaki, M., Yamashita, K.: A new approach to enhancement of bone formation by electrically polarized hydroxyapatite, *J. Dent. Res.*, **80**, 1925–1929, (2001).
- 94 Kobayashi, T., Nakamura, S., Yamashita, K.: Enhanced osteobonding by negative surface charges of electrically polarized hydroxyapatite, *J. Biomed. Mater. Res.*, **57**, 477–484, (2001).
- 95 Park, Y.J., Hwang, K.S., Song, J.E., Ong, J.L., Rawls, H.R.: Growth of calcium phosphate on poling treated ferroelectric BaTiO₃ ceramics, *Biomaterials*, **23**, 3859–3864, (2002).
- 96 Hwang, K.S., Song, J.E., Jo, J.W., Yang, H.S., Park, Y.J., Ong, J.L., Rawls, H.R.: Effect of poling conditions on growth of calcium phosphate crystal in ferroelectric BaTiO₃ ceramics, *J. Mater. Sci. Mater. Med.*, **13**, 133–138, (2002).
- 97 Yamashita, K.: Enhanced bioactivity of electrically poled hydroxyapatite ceramics and coatings, *Mater. Sci. Forum*, **426–432**, 3237–3242, (2003).
- 98 Nakamura, S., Kobayashi, T., Yamashita, K.: Highly orientated calcification in newly formed bones on negatively charged hydroxyapatite electrets, *Key Eng. Mater.*, **284–286**, 897–900, (2005).
- 99 Kato, R., Nakamura, S., Katayama, K., Yamashita, K.: Electrical polarization of plasma-spray-hydroxyapatite coatings for improvement of osteoconduction of implants, *J. Biomed. Mater. Res. A*, **74A**, 652–658, (2005).
- 100 Nakamura, S., Kobayashi, T., Nakamura, M., Itoh, S., Yamashita, K.: Electrostatic surface charge acceleration of bone ingrowth of porous hydroxyapatite/ β -tricalcium phosphate ceramics, *J. Biomed. Mater. Res. A*, **92A**, 267–275, (2010).
- 101 Tarafder, S., Bodhak, S., Bandyopadhyay, A., Bose, S.: Effect of electrical polarization and composition of biphasic calcium phosphates on early stage osteoblast interactions, *J. Biomed. Mater. Res. B Appl. Biomater.*, **97B**, 306–314, (2011).
- 102 Ohba, S., Wang, W., Itoh, S., Takagi, Y., Nagai, A., Yamashita, K.: Acceleration of new bone formation by an electrically polarized hydroxyapatite microgranule/platelet-rich plasma composite, *Acta Biomater.*, **8**, 2778–2787, (2012).
- 103 Tarafder, S., Banerjee, S., Bandyopadhyay, A., Bose, S.: Electrically polarized biphasic calcium phosphates: adsorption and release of bovine serum albumin, *Langmuir*, **26**, 16625–16629, (2010).

- ¹⁰⁴ Itoh, S., Nakamura, S., Nakamura, M., Shinomiya, K., Yamashita, K.: Enhanced bone regeneration by electrical polarization of hydroxyapatite, *Artif. Organs*, **30**, 863–869, (2006).
- ¹⁰⁵ Nakamura, M., Nagai, A., Ohashi, N., Tanaka, Y., Sekilima, Y., Nakamura, S.: Regulation of osteoblast-like cell behaviors on hydroxyapatite by electrical polarization, *Key Eng. Mater.*, **361–363**, 1055–1058, (2008).
- ¹⁰⁶ Nakamura, M., Nagai, A., Tanaka, Y., Sekilima, Y., Yamashita, K.: Polarized hydroxyapatite promotes spread and motility of osteoblastic cells, *J. Biomed. Mater. Res. A*, **92A**, 783–790, (2010).
- ¹⁰⁷ Nakamura, M., Nagai, A., Yamashita, K.: Surface electric fields of apatite electret promote osteoblastic responses, *Key Eng. Mater.*, **529–530**, 357–360, (2013).
- ¹⁰⁸ Nakamura, S., Kobayashi, T., Yamashita, K.: Extended bioactivity in the proximity of hydroxyapatite ceramic surfaces induced by polarization charges, *J. Biomed. Mater. Res.*, **61**, 593–599, (2002).
- ¹⁰⁹ Wang, W., Itoh, S., Tanaka, Y., Nagai, A., Yamashita, K.: Comparison of enhancement of bone ingrowth into hydroxyapatite ceramics with highly and poorly interconnected pores by electrical polarization, *Acta Biomater.*, **5**, 3132–3140, (2009).
- ¹¹⁰ Cartmell, S.H., Thurstan, S., Gittings, J.P., Griffiths, S., Bowen, C.R., Turner, I.G.: Polarization of porous hydroxyapatite scaffolds: influence on osteoblast cell proliferation and extracellular matrix production, *J. Biomed. Mater. Res. A*, **102A**, 1047–1052, (2014).
- ¹¹¹ Nakamura, M., Kobayashi, A., Nozaki, K., Horiuchi, N., Nagai, A., Yamashita, K.: Improvement of osteoblast adhesion through polarization of plasma-sprayed hydroxyapatite coatings on metal, *J. Med. Biol. Eng.*, **34**, 44–48, (2014).
- ¹¹² Ioku, K.: Tailored bioceramics of calcium phosphates for regenerative medicine, *J. Ceram. Soc. Jpn.*, **118**, 775–783, (2010).
- ¹¹³ Fang, Y., Agrawal, D.K., Roy, D.M., Roy, R.: Fabrication of transparent hydroxyapatite ceramics by ambient-pressure sintering, *Mater. Lett.*, **23**, 147–151, (1995).
- ¹¹⁴ Varma, H., Vijayan, S.P., Babu, S.S.: Transparent hydroxyapatite ceramics through gel-casting and low-temperature sintering, *J. Am. Ceram. Soc.*, **85**, 493–495, (2002).
- ¹¹⁵ Watanabe, Y., Ikoma, T., Monkawa, A., Suetsugu, Y., Yamada, H., Tanaka, J., Moriyoshi, Y.: Fabrication of transparent hydroxyapatite sintered body with high crystal orientation by pulse electric current sintering, *J. Am. Ceram. Soc.*, **88**, 243–245, (2005).
- ¹¹⁶ Kotobuki, N., Ioku, K., Kawagoe, D., Fujimori, H., Goto, S., Ohgushi, H.: Observation of osteogenic differentiation cascade of living mesenchymal stem cells on transparent hydroxyapatite ceramics, *Biomaterials*, **26**, 779–785, (2005).
- ¹¹⁷ John, A., Varma, H.K., Vijayan, S., Bernhardt, A., Lode, A., Vogel, A., Burmeister, B., Hanke, T., Domaschke, H., Gelinisky, M.: In vitro investigations of bone remodeling on a transparent hydroxyapatite ceramic, *Biomed. Mater.*, **4**, 015007, (2009).
- ¹¹⁸ Wang, J., Shaw, L.L.: Transparent nanocrystalline hydroxyapatite by pressure-assisted sintering, *Scripta Mater.*, **63**, 593–596, (2010).
- ¹¹⁹ Tan, N., Kou, Z., Ding, Y., Leng, Y., Liu, C., He, D.: Novel substantial reductions in sintering temperatures for preparation of transparent hydroxyapatite bioceramics under ultra-high pressure, *Scripta Mater.*, **65**, 819–822, (2011).
- ¹²⁰ Boilet, L., Descamps, M., Rguiti, E., Tricoteaux, A., Lu, J., Petit, F., Lardot, V., Cambier, F., Leriche, A.: Processing and properties of transparent hydroxyapatite and β tricalcium phosphate obtained by HIP process, *Ceram. Int.*, **39**, 283–288, (2013).
- ¹²¹ Han, Y.H., Kim, B.N., Yoshida, H., Yun, J., Son, H.W., Lee, J., Kim, S.: Spark plasma sintered superplastic deformed transparent ultrafine hydroxyapatite nanoceramics, *Adv. Appl. Ceram.*, **115**, 174–184, (2016).
- ¹²² Chesnaud, A., Bogicevic, C., Karolak, F., Estournès, C., Dezanneau, G.: Preparation of transparent oxyapatite ceramics by combined use of freeze-drying and spark-plasma sintering, *Chem. Comm.*, 1550–1552, (2007).
- ¹²³ Eriksson, M., Liu, Y., Hu, J., Gao, L., Nygren, M., Shen, Z.: Transparent hydroxyapatite ceramics with nanograin structure prepared by high pressure spark plasma sintering at the minimized sintering temperature, *J. Eur. Ceram. Soc.*, **31**, 1533–1540, (2011).
- ¹²⁴ Liu, Y., Shen, Z.: Dehydroxylation of hydroxyapatite in dense bulk ceramics sintered by spark plasma sintering, *J. Eur. Ceram. Soc.*, **32**, 2691–2696, (2012).
- ¹²⁵ Yoshida, H., Kim, B.N., Son, H.W., Han, Y.H., Kim, S.: Superplastic deformation of transparent hydroxyapatite, *Scripta Mater.*, **69**, 155–158, (2013).
- ¹²⁶ Kim, B.N., Prajatelista, E., Han, Y.H., Son, H.W., Sakka, Y., Kim, S.: Transparent hydroxyapatite ceramics consolidated by spark plasma sintering, *Scripta Mater.*, **69**, 366–369, (2013).
- ¹²⁷ Yun, J., Son, H., Prajatelista, E., Han, Y.H., Kim, S., Kim, B.N.: Characterisation of transparent hydroxyapatite nanoceramics prepared by spark plasma sintering, *Adv. Appl. Ceram.*, **113**, 67–72, (2014).
- ¹²⁸ Li, Z., Khor, K.A.: Transparent hydroxyapatite obtained through spark plasma sintering: optical and mechanical properties, *Key Eng. Mater.*, **631**, 51–56, (2015).
- ¹²⁹ Kobune, M., Mineshige, A., Fujii, S., Iida, H.: Preparation of translucent hydroxyapatite ceramics by HIP and their physical properties, *J. Ceram. Soc. Jpn.*, **105**, 210–213, (1997).
- ¹³⁰ Barralet, J.E., Fleming, G.J.P., Campion, C., Harris, J.J., Wright, A.J.: Formation of translucent hydroxyapatite ceramics by sintering in carbon dioxide atmospheres, *J. Mater. Sci.*, **38**, 3979–3993, (2003).
- ¹³¹ Chaudhry, A.A., Yan, H., Gong, K., Inam, F., Viola, G., Reece, M.J., Goodall, J.B.M., ur Rehman, I., McNeil-Watson, F.K., Corbett, J.C.W., Knowles, J.C., Darr, J.A.: High-strength nanograined and translucent hydroxyapatite monoliths via continuous hydrothermal synthesis and optimized spark plasma sintering, *Acta Biomater.*, **7**, 791–799, (2011).
- ¹³² Tancred, D.C., McCormack, B.A., Carr, A.J.: A synthetic bone implant macroscopically identical to cancellous bone, *Biomaterials*, **19**, 2303–2311, (1998).
- ¹³³ Miao, X., Sun, D.: Graded/gradient porous biomaterials, *Materials*, **3**, 26–47, (2010).
- ¹³⁴ Schliephake, H., Neukam, F.W., Klosa, D.: Influence of pore dimensions on bone ingrowth into porous hydroxyapatite blocks used as bone graft substitutes. A histometric study, *Int. J. Oral Maxillofac. Surg.*, **20**, 53–58, (1991).
- ¹³⁵ Otsuki, B., Takemoto, M., Fujibayashi, S., Neo, M., Kokubo, T., Nakamura, T.: Pore throat size and connectivity determine bone and tissue ingrowth into porous implants: three-dimensional micro-CT based structural analyses of porous bioactive titanium implants, *Biomaterials*, **27**, 5892–5900, (2006).
- ¹³⁶ Hing, K.A., Best, S.M., Bonfield, W.: Characterization of porous hydroxyapatite, *J. Mater. Sci. Mater. Med.*, **10**, 135–145, (1999).
- ¹³⁷ Lu, J.X., Flautre, B., Anselme, K., Hardouin, P., Gallur, A., Descamps, M., Thierry, B.: Role of interconnections in porous bioceramics on bone recolonization in vitro and in vivo, *J. Mater. Sci. Mater. Med.*, **10**, 111–120, (1999).
- ¹³⁸ Karageorgiou, V., Kaplan, D.: Porosity of 3D biomaterial scaffolds and osteogenesis, *Biomaterials*, **26**, 5474–5491, (2005).
- ¹³⁹ Jones, A.C., Arns, C.H., Sheppard, A.P., Hutmacher, D.W., Milthorpe, B.K., Knackstedt, M.A.: Assessment of bone ingrowth into porous biomaterials using MICRO-CT, *Biomaterials*, **28**, 2491–2504, (2007).

- 140 Tamai, N., Myoui, A., Kudawara, I., Ueda, T., Yoshikawa, H.: Novel fully interconnected porous hydroxyapatite ceramic in surgical treatment of benign bone tumor, *J. Orthop. Sci.*, **15**, 560–568, (2010).
- 141 Sakane, M., Tsukanishi, T., Funayama, T., Kobayashi, M., Ochiai, N.: Unidirectional porous β -tricalcium phosphate bone substitute: examination of balance between new bone formation and absorption, *Key Eng. Mater.*, **493–494**, 132–134, (2012).
- 142 Panzavolta, S., Torricelli, P., Amadori, S., Parrilli, A., Rubini, K., Della Bella, E., Fini, M., Bigi, A.: 3D interconnected porous biomimetic scaffolds: in vitro cell response, *J. Biomed. Mater. Res. A*, **101A**, 3560–3570, (2013).
- 143 Jin, L., Feng, Z.Q., Wang, T., Ren, Z., Ma, S., Wu, J., Sun, D.: A novel fluffy hydroxylapatite fiber scaffold with deep interconnected pores designed for three-dimensional cell culture, *J. Mater. Chem. B*, **2**, 129–136, (2014).
- 144 Flautre, B., Descamps, M., Delecourt, C., Blary, M.C., Hardouin, P.: Porous HA ceramic for bone replacement: role of the pores and interconnections-experimental study in the rabbits, *J. Mater. Sci. Mater. Med.*, **12**, 679–682, (2001).
- 145 Tamai, N., Myoui, A., Tomita, T., Nakase, T., Tanaka, J., Ochi, T., Yoshikawa, H.: Novel hydroxyapatite ceramics with an interconnective porous structure exhibit superior osteoconduction in vivo, *J. Biomed. Mater. Res.*, **59**, 110–117, (2002).
- 146 Mastrogiacomo, M., Scaglione, S., Martinetti, R., Dolcini, L., Beltrame, F., Cancedda, R., Quarto, R.: Role of scaffold internal structure on in vivo bone formation in macroporous calcium phosphate bioceramics, *Biomaterials*, **27**, 3230–3237, (2006).
- 147 Okamoto, M., Dohi, Y., Ohgushi, H., Shimaoka, H., Ikeuchi, M., Matsushima, A., Yonemasu, K., Hosoi, H.: Influence of the porosity of hydroxyapatite ceramics on in vitro and in vivo bone formation by cultured rat bone marrow stromal cells. *J. Mater. Sci. Mater. Med.*, **17**, 327–336, (2006).
- 148 Zhang, L., Hanagata, N., Maeda, M., Minowa, T., Ikoma, T., Fan, H., Zhang, X.: Porous hydroxyapatite and biphasic calcium phosphate ceramics promote ectopic osteoblast differentiation from mesenchymal stem cells, *Sci. Technol. Adv. Mater.*, **10**, 025003, (2009).
- 149 Li, X., Liu, H., Niu, X., Fan, Y., Feng, Q., Cui, F.Z., Watari, F.: Osteogenic differentiation of human adipose-derived stem cells induced by osteoinductive calcium phosphate ceramics, *J. Biomed. Mater. Res. B Appl. Biomater.*, **97B**, 10–19, (2011).
- 150 Hong, M.H., Kim, S.M., Han, M.H., Kim, Y.H., Lee, Y.K., Oh, D.S.: Evaluation of microstructure effect of the porous spherical β -tricalcium phosphate granules on cellular responses, *Ceram. Int.*, **40**, 6095–6102, (2014).
- 151 de Godoy, R.F., Hutchens, S., Campion, C., Blunn, G.: Silicate-substituted calcium phosphate with enhanced strut porosity stimulates osteogenic differentiation of human mesenchymal stem cells, *J. Mater. Sci. Mater. Med.*, **26**, 54, (2015).
- 152 Omae, H., Mochizuki, Y., Yokoya, S., Adachi, N., Ochi, M.: Effects of interconnecting porous structure of hydroxyapatite ceramics on interface between grafted tendon and ceramics, *J. Biomed. Mater. Res. A*, **79A**, 329–337, (2006).
- 153 Yoshikawa, H., Tamai, N., Murase, T., Myoui, A.: Interconnected porous hydroxyapatite ceramics for bone tissue engineering, *J. R. Soc. Interface*, **6**, S341–S348, (2009).
- 154 Ribeiro, G.B.M., Trommer, R.M., dos Santos, L.A., Bergmann, C.P.: Novel method to produce β -TCP scaffolds, *Mater. Lett.*, **65**, 275–277, (2011).
- 155 Silva, T.S.N., Primo, B.T., Silva, Jr., A.N., Machado, D.C., Viezzer, C., Santos, L.A.: Use of calcium phosphate cement scaffolds for bone tissue engineering: in vitro study, *Acta Cir. Bras.*, **26**, 7–11, (2011).
- 156 de Moraes MacHado, J.L., Giehl, I.C., Nardi, N.B., dos Santos, L.A.: Evaluation of scaffolds based on α -tricalcium phosphate cements for tissue engineering applications, *IEEE Trans. Biomed. Eng.*, **58**, 1814–1819, (2011).
- 157 Li, S.H., de Wijn, J.R., Layrolle, P., de Groot, K.: Novel method to manufacture porous hydroxyapatite by dual-phase mixing, *J. Am. Ceram. Soc.*, **86**, 65–72, (2003).
- 158 de Oliveira, J.F., de Aguiar, P.F., Rossi, A.M., Soares, G.D.A.: Effect of process parameters on the characteristics of porous calcium phosphate ceramics for bone tissue scaffolds, *Artif. Organs*, **27**, 406–411, (2003).
- 159 Swain, S.K., Bhattacharyya, S.: Preparation of high strength macroporous hydroxyapatite scaffold, *Mater. Sci. Eng. C*, **33**, 67–71, (2013).
- 160 Maeda, H., Kasuga, T., Nogami, M., Kagami, H., Hata, K., Ueda, M.: Preparation of bonelike apatite composite sponge, *Key Eng. Mater.*, **254–256**, 497–500, (2004).
- 161 le Ray, A.M., Gautier, H., Bouler, J.M., Weiss, P., Merle, C.: A new technological procedure using sucrose as porogen compound to manufacture porous biphasic calcium phosphate ceramics of appropriate micro- and macrostructure, *Ceram. Int.*, **36**, 93–101, (2010).
- 162 Li, S.H., de Wijn J.R., Layrolle, P., de Groot, K.: Synthesis of macroporous hydroxyapatite scaffolds for bone tissue engineering, *J. Biomed. Mater. Res.*, **61**, 109–120, (2002).
- 163 Hesaraki, S., Sharifi, D.: Investigation of an effervescent additive as porogenic agent for bone cement macroporosity, *Biomed. Mater. Eng.*, **17**, 29–38, (2007).
- 164 Hesaraki, S., Moztafzadeh, F., Sharifi, D.: Formation of interconnected macropores in apatitic calcium phosphate bone cement with the use of an effervescent additive, *J. Biomed. Mater. Res. A*, **83A**, 80–87, (2007).
- 165 Pal, K., Pal, S.: Development of porous hydroxyapatite scaffolds, *Mater. Manuf. Process.*, **21**, 325–328, (2006).
- 166 Tas, A.C.: Preparation of porous apatite granules from calcium phosphate cement, *J. Mater. Sci. Mater. Med.*, **19**, 2231–2239, (2008).
- 167 Yao, X., Tan, S., Jiang, D.: Improving the properties of porous hydroxyapatite ceramics by fabricating methods, *J. Mater. Sci.*, **40**, 4939–4942, (2005).
- 168 Song, H.Y., Youn, M.H., Kim, Y.H., Min, Y.K., Yang, H.M., Lee, B.T.: Fabrication of porous β -TCP bone graft substitutes using PMMA powder and their biocompatibility study, *Korean J. Mater. Res.*, **17**, 318–322, (2007).
- 169 Youn, M.H., Paul, R.K., Song, H.Y., Lee, B.T.: Fabrication of porous structure of BCP sintered bodies using microwave assisted synthesized HAp nano powder, *Mater. Sci. Forum*, **534–536**, 49–52, (2007).
- 170 Almirall, A., Larrecq, G., Delgado, J.A., Martínez, S., Ginebra, M.P., Planell, J.A.: Fabrication of low temperature hydroxyapatite foams. *Key Eng. Mater.* **254–256**, 1001–1004, (2004).
- 171 Almirall, A., Larrecq, G., Delgado, J.A., Martínez, S., Planell, J.A., Ginebra, M.P.: Fabrication of low temperature macroporous hydroxyapatite scaffolds by foaming and hydrolysis of an α -TCP paste, *Biomaterials*, **25**, 3671–3680, (2004).
- 172 Huang, X., Miao, X.: Novel porous hydroxyapatite prepared by combining H_2O_2 foaming with PU sponge and modified with PLGA and bioactive glass, *J. Biomater. Appl.*, **21**, 351–374, (2007).
- 173 Strnadova, M., Protivinsky, J., Strnad, J., Vejsicka, Z.: Preparation of porous synthetic nanostructured HA scaffold, *Key Eng. Mater.*, **361–363**, 211–214, (2008).
- 174 Li, B., Chen, X., Guo, B., Wang, X., Fan, H., Zhang, X.: Fabrication and cellular biocompatibility of porous carbonated biphasic calcium phosphate ceramics with a nanostructure, *Acta Biomater.*, **5**, 134–143, (2009).
- 175 Cheng, Z., Zhao, K., Wu, Z.P.: Structure control of hydroxyapatite ceramics via an electric field assisted freeze casting method, *Ceram. Int.*, **41**, 8599–8604, (2015).

- 176 Takagi, S., Chow, L.C.: Formation of macropores in calcium phosphate cement implants, *J. Biomed. Mater. Res.*, **12**, 135–139, (2001).
- 177 Walsh, D., Tanaka, J.: Preparation of a bone-like apatite foam cement, *J. Mater. Sci. Mater. Med.*, **12**, 339–344, (2001).
- 178 Tadic, D., Beckmann, F., Schwarz, K., Epple, M.: A novel method to produce hydroxylapatite objects with interconnecting porosity that avoids sintering, *Biomaterials*, **25**, 3335–3340, (2004).
- 179 Komlev, V.S., Barinov, S.M.: Porous hydroxyapatite ceramics of bi-modal pore size distribution, *J. Mater. Sci. Mater. Med.*, **13**, 295–299, (2002).
- 180 Sepulveda, P., Binner, J.G., Rogero, S.O., Higa, O.Z., Bressiani, J.C.: Production of porous hydroxyapatite by the gel-casting of foams and cytotoxic evaluation, *J. Biomed. Mater. Res.*, **50**, 27–34, (2000).
- 181 Hsu, Y.H., Turner, I.G., Miles, A.W.: Mechanical characterization of dense calcium phosphate bioceramics with interconnected porosity, *J. Mater. Sci. Mater. Med.*, **18**, 2319–2329, (2007).
- 182 Zhang, H.G., Zhu, Q.: Preparation of porous hydroxyapatite with interconnected pore architecture, *J. Mater. Sci. Mater. Med.*, **18**, 1825–1829, (2007).
- 183 Chevalier, E., Chulia, D., Pouget, C., Viana, M.: Fabrication of porous substrates: a review of processes using pore forming agents in the biomaterial field, *J. Pharm. Sci.*, **97**, 1135–1154, (2008).
- 184 Tang, Y.J., Tang, Y.F., Lv, C.T., Zhou, Z.H.: Preparation of uniform porous hydroxyapatite biomaterials by a new method, *Appl. Surf. Sci.*, **254**, 5359–5362, (2008).
- 185 Abdulqader, S.T., Rahman, I.A., Ismail, H., Ponnuraj Kannan, T., Mahmood, Z.: A simple pathway in preparation of controlled porosity of biphasic calcium phosphate scaffold for dentin regeneration, *Ceram. Int.*, **39**, 2375–2381, (2013).
- 186 Stares, S.L., Fredel, M.C., Greil, P., Travitzky, N.: Paper-derived hydroxyapatite, *Ceram. Int.*, **39**, 7179–7183, (2013).
- 187 Wen, F.H., Wang, F., Gai, Y., Wang, M.T., Lai, Q.H.: Preparation of mesoporous hydroxylapatite ceramics using polystyrene microspheres as template, *Appl. Mech. Mater.*, **389**, 194–198, (2013).
- 188 Guda, T., Appleford, M., Oh, S., Ong, J.L.: A cellular perspective to bioceramic scaffolds for bone tissue engineering: the state of the art, *Curr. Top. Med. Chem.*, **8**, 290–299, (2008).
- 189 Habraken, W.J.E.M., Wolke, J.G.C., Jansen, J.A.: Ceramic composites as matrices and scaffolds for drug delivery in tissue engineering, *Adv. Drug Deliv. Rev.*, **59**, 234–248, (2007).
- 190 Tian, J., Tian, J.: Preparation of porous hydroxyapatite, *J. Mater. Sci.*, **36**, 3061–3066, (2001).
- 191 Swain, S.K., Bhattacharyya, S., Sarkar, D.: Preparation of porous scaffold from hydroxyapatite powders, *Mater. Sci. Eng. C*, **31**, 1240–1244, (2011).
- 192 Zhao, K., Tang, Y.F., Qin, Y.S., Luo, D.F.: Polymer template fabrication of porous hydroxyapatite scaffolds with interconnected spherical pores, *J. Eur. Ceram. Soc.*, **31**, 225–229, (2011).
- 193 Sung, J.H., Shin, K.H., Moon, Y.W., Koh, Y.H., Choi, W.Y., Kim, H.E.: Production of porous calcium phosphate (CaP) ceramics with highly elongated pores using carbon-coated polymeric templates, *Ceram. Int.*, **38**, 93–97, (2012).
- 194 Oha, D.S., Kim, Y.H., Ganbat, D., Han, M.H., Lim, P., Back, J.H., Lee, F.Y., Tawfeek, H.: Bone marrow absorption and retention properties of engineered scaffolds with micro-channels and nano-pores for tissue engineering: a proof of concept, *Ceram. Int.*, **39**, 8401–8410, (2013).
- 195 Deville, S., Saiz, E., Tomsia, A.P.: Freeze casting of hydroxyapatite scaffolds for bone tissue engineering, *Biomaterials*, **27**, 5480–5489, (2006).
- 196 Lee, E.J., Koh, Y.H., Yoon, B.H., Kim, H.E., Kim, H.W.: Highly porous hydroxyapatite bioceramics with interconnected pore channels using camphene-based freeze casting, *Mater. Lett.*, **61**, 2270–2273, (2007).
- 197 Fu, Q., Rahaman, M.N., Dogan, F., Bal, B.S.: Freeze casting of porous hydroxyapatite scaffolds. I. processing and general microstructure, *J. Biomed. Mater. Res. B Appl. Biomater.*, **86B**, 125–135, (2008).
- 198 Impens, S., Schelstraete, R., Luyten, J., Mullens, S., Thijs, I., van Humbeeck, J., Schrooten, J.: Production and characterisation of porous calcium phosphate structures with controllable hydroxyapatite/ β -tricalcium phosphate ratios, *Adv. Appl. Ceram.*, **108**, 494–500, (2009).
- 199 Macchetta, A., Turner, I.G., Bowen, C.R.: Fabrication of HA/TCP scaffolds with a graded and porous structure using a camphene-based freeze-casting method, *Acta Biomater.*, **5**, 1319–1327, (2009).
- 200 Potoczek, M., Zima, A., Paszkiewicz, Z., Ślósarczyk, A.: Manufacturing of highly porous calcium phosphate bioceramics via gel-casting using agarose, *Ceram. Int.*, **35**, 2249–2254, (2009).
- 201 Zuo, K.H., Zeng, Y.P., Jiang, D.: Effect of polyvinyl alcohol additive on the pore structure and morphology of the freeze-cast hydroxyapatite ceramics, *Mater. Sci. Eng. C*, **30**, 283–287, (2010).
- 202 Soon, Y.M., Shin, K.H., Koh, Y.H., Lee, J.H., Choi, W.Y., Kim, H.E.: Fabrication and compressive strength of porous hydroxyapatite scaffolds with a functionally graded core/shell structure, *J. Eur. Ceram. Soc.*, **31**, 13–18, (2011).
- 203 Hesaraki, S.: Freeze-casted nanostructured apatite scaffold obtained from low temperature biomineralization of reactive calcium phosphates, *Key Eng. Mater.*, **587**, 21–26, (2014).
- 204 Ng, S., Guo, J., Ma, J., Loo, S.C.J.: Synthesis of high surface area mesostructured calcium phosphate particles, *Acta Biomater.*, **6**, 3772–3781, (2010).
- 205 Walsh, D., Hopwood, J.D., Mann, S.: Crystal tectonics: construction of reticulated calcium phosphate frameworks in bicontinuous reverse microemulsions, *Science*, **264**, 1576–1578, (1994).
- 206 Walsh, D., Mann, S.: Chemical synthesis of microskeletal calcium phosphate in bicontinuous microemulsions, *Chem. Mater.*, **8**, 1944–1953, (1996).
- 207 Zhao, K., Tang, Y.F., Qin, Y.S., Wei, J.Q.: Porous hydroxyapatite ceramics by ice templating: freezing characteristics and mechanical properties, *Ceram. Int.*, **37**, 635–639, (2011).
- 208 Zhou, K., Zhang, Y., Zhang, D., Zhang, X., Li, Z., Liu, G., Button, T.W.: Porous hydroxyapatite ceramics fabricated by an ice-templating method, *Scripta Mater.*, **64**, 426–429, (2011).
- 209 Flauder, S., Gbureck, U., Muller, F.A.: TCP scaffolds with an interconnected and aligned porosity fabricated via ice-templating, *Key Eng. Mater.*, **529–530**, 129–132, (2013).
- 210 Zhang, Y., Zhou, K., Bao, Y., Zhang, D.: Effects of rheological properties on ice-templated porous hydroxyapatite ceramics, *Mater. Sci. Eng. C*, **33**, 340–346, (2013).
- 211 White, E., Shors, E.C.: Biomaterial aspects of Interpore-200 porous hydroxyapatite, *Dent. Clin. North Am.*, **30**, 49–67, (1986).
- 212 Aizawa, M., Howell, S.F., Itatani, K., Yokogawa, Y., Nishizawa, K., Toriyama, M., Kameyama, T.: Fabrication of porous ceramics with well-controlled open pores by sintering of fibrous hydroxyapatite particles, *J. Ceram. Soc. Jpn.*, **108**, 249–253, (2000).
- 213 Nakahira, A., Tamai, M., Sakamoto, K., Yamaguchi, S.: Sintering and microstructure of porous hydroxyapatite, *J. Ceram. Soc. Jpn.*, **108**, 99–104, (2000).
- 214 Rodriguez-Lorenzo, L.M., Vallet-Regí, M., Ferreira, J.M.F.: Fabrication of porous hydroxyapatite bodies by a new direct

- consolidation method: starch consolidation, *J. Biomed. Mater. Res.*, **60**, 232–240, (2002).
- 215 Charriere, E., Lemaitre, J., Zysset, P.: Hydroxyapatite cement scaffolds with controlled macroporosity: fabrication protocol and mechanical properties, *Biomaterials*, **24**, 809–817, (2003).
- 216 Eichenseer, C., Will, J., Rampf, M., Wend, S., Greil, P.: Biomorphous porous hydroxyapatite-ceramics from rattan (*Calamus Rotang*), *J. Mater. Sci. Mater. Med.*, **21**, 131–137, (2010).
- 217 Zhou, L., Wang, D., Huang, W., Yao, A., Kamitakahara, M., Ioku, K.: Preparation and characterization of periodic porous frame of hydroxyapatite, *J. Ceram. Soc. Jpn.*, **117**, 521–524, (2009).
- 218 Kawata, M., Uchida, H., Itatani, K., Okada, I., Koda, S., Aizawa, M.: Development of porous ceramics with well-controlled porosities and pore sizes from apatite fibers and their evaluations, *J. Mater. Sci. Mater. Med.*, **15**, 817–823, (2004).
- 219 Koh, Y.H., Kim, H.W., Kim, H.E., Halloran, J.W.: Fabrication of macrochannelled-hydroxyapatite bioceramic by a coextrusion process, *J. Am. Ceram. Soc.*, **85**, 2578–2580, (2002).
- 220 Kitamura, M., Ohtsuki, C., Ogata, S.I., Kamitakahara, M., Tanihara, M., Miyazaki, T.: Mesoporous calcium phosphate via post-treatment of α -TCP, *J. Am. Ceram. Soc.*, **88**, 822–826, (2005).
- 221 Walsh, D., Boanini, E., Tanaka, J., Mann, S.: Synthesis of tri-calcium phosphate sponges by interfacial deposition and thermal transformation of self-supporting calcium phosphate films, *J. Mater. Chem.*, **15**, 1043–1048, (2005).
- 222 Gonzalez-McQuire, R., Green, D., Walsh, D., Hall, S., Chan-Ching, J.Y., Oreffo, R.O.C., Mann, S.: Fabrication of hydroxyapatite sponges by dextran sulphate/amino acid templating, *Biomaterials*, **26**, 6652–6656, (2005).
- 223 Xu, S., Li, D., Lu, B., Tang, Y., Wang, C., Wang, Z.: Fabrication of a calcium phosphate scaffold with a three dimensional channel network and its application to perfusion culture of stem cells, *Rapid Prototyping J.*, **13**, 99–106, (2007).
- 224 Saiz, E., Gremillard, L., Menendez, G., Miranda, P., Gryn, K., Tomsia, A.P.: Preparation of porous hydroxyapatite scaffolds, *Mater. Sci. Eng. C*, **27**, 546–550, (2007).
- 225 Kamitakahara, M., Ohtsuki, C., Kawachi, G., Wang, D., Ioku, K.: Preparation of hydroxyapatite porous ceramics with different porous structures using a hydrothermal treatment with different aqueous solutions, *J. Ceram. Soc. Jpn.*, **116**, 6–9, (2008).
- 226 Peña, J., Román, J., Cabañas, M.V., Vallet-Regí, M.: An alternative technique to shape scaffolds with hierarchical porosity at physiological temperature, *Acta Biomater.*, **6**, 1288–1296, (2010).
- 227 Nakamura, S., Nakahira, A.: Synthesis and evaluation of porous hydroxyapatite prepared by hydrothermal treatment and subsequent sintering method, *J. Ceram. Soc. Jpn.*, **116**, 42–45, (2008).
- 228 Zhang, J., Fujiwara, M., Xu, Q., Zhu, Y., Iwasa, M., Jiang, D.: Synthesis of mesoporous calcium phosphate using hybrid templates, *Micropor. Mesopor. Mater.*, **111**, 411–416, (2008).
- 229 Song, H.Y., Islam, S., Lee, B.T.: A novel method to fabricate unidirectional porous hydroxyapatite body using ethanol bubbles in a viscous slurry, *J. Am. Ceram. Soc.*, **91**, 3125–3127, (2008).
- 230 Kawachi, G., Misumi, H., Fujimori, H., Goto, S., Ohtsuki, C., Kamitakahara, M., Ioku, K.: Fabrication of porous blocks of calcium phosphate through hydrothermal processing under glycine coexistence, *J. Ceram. Soc. Jpn.*, **118**, 559–563, (2010).
- 231 Sakamoto, M., Nakasu, M., Matsumoto, T., Okihana, H.: Development of superporous hydroxyapatites and their examination with a culture of primary rat osteoblasts, *J. Biomed. Mater. Res. A*, **82A**, 238–242, (2007).
- 232 Sakamoto, M.: Development and evaluation of superporous hydroxyapatite ceramics with triple pore structure as bone tissue scaffold, *J. Ceram. Soc. Jpn.*, **118**, 753–757, (2010).
- 233 Sakamoto, M., Matsumoto, T.: Development and evaluation of superporous ceramics bone tissue scaffold materials with triple pore structure a) hydroxyapatite, b) beta-tricalcium phosphate. In: Bone regeneration. Tal, H. (Ed.). InTech Europe, Rijeka, Croatia, 301–320, (2012).
- 234 Deisinger, U.: Generating porous ceramic scaffolds: processing and properties, *Key Eng. Mater.*, **441**, 155–179, (2010).
- 235 Ishikawa, K., Tsuru, K., Pham, T.K., Maruta, M., Matsuya, S.: Fully-interconnected pore forming calcium phosphate cement, *Key Eng. Mater.*, **493–494**, 832–835, (2012).
- 236 Yoon, H.J., Kim, U.C., Kim, J.H., Koh, Y.H., Choi, W.Y., Kim, H.E.: Fabrication and characterization of highly porous calcium phosphate (CaP) ceramics by freezing foamed aqueous CaP suspensions, *J. Ceram. Soc. Jpn.*, **119**, 573–576, (2011).
- 237 Ahn, M.K., Shin, K.H., Moon, Y.W., Koh, Y.H., Choi, W.Y., Kim, H.E.: Highly porous biphasic calcium phosphate (BCP) ceramics with large interconnected pores by freezing vigorously foamed BCP suspensions under reduced pressure, *J. Am. Ceram. Soc.*, **94**, 4154–4156, (2011).
- 238 Ji, L., Jell, G., Dong, Y., Jones, J.R., Stevens, M.M.: Template synthesis of ordered macroporous hydroxyapatite bioceramics, *Chem. Commun.*, **47**, 9048–9050, (2011).
- 239 Wang, X.Y., Han, Y.C., Li, S.P.: Preparation and characterization of calcium phosphate crystals by precursor thermolysis method, *Key Eng. Mater.*, **493–494**, 191–194, (2012).
- 240 Schlosser, M., Kleebe, H.J.: Vapor transport sintering of porous calcium phosphate ceramics, *J. Am. Ceram. Soc.*, **95**, 1581–1587, (2012).
- 241 Tanaka, T., Yoshioka, T., Ikoma, T., Kuwayama, T., Higaki, T., Tanaka, M.: Fabrication of three different types of porous carbonate-substituted apatite ceramics for artificial bone, *Key Eng. Mater.*, **529–530**, 143–146, (2013).
- 242 Zheng, W., Liu, G., Yan, C., Xiao, Y., Miao, X.G.: Strong and bioactive tri-calcium phosphate scaffolds with tube-like macropores, *J. Biomim. Biomater. Tissue Eng.*, **19**, 65–75, (2014).
- 243 Tsuru, K., Nikaido, T., Munar, M.L., Maruta, M., Matsuya, S., Nakamura, S., Ishikawa, K.: Synthesis of carbonate apatite foam using β -TCP foams as precursors, *Key Eng. Mater.*, **587**, 52–55, (2014).
- 244 Chen, Z.C., Zhang, X.L., Zhou, K., Cai, H., Liu, C.Q.: Novel fabrication of hierarchically porous hydroxyapatite scaffolds with refined porosity and suitable strength, *Adv. Appl. Ceram.*, **114**, 183–187, (2015).
- 245 Swain, S.K., Bhattacharyya, S., Sarkar, D.: Fabrication of porous hydroxyapatite scaffold via polyethylene glycol-polyvinyl alcohol hydrogel state, *Mater. Res. Bull.*, **64**, 257–261, (2015).
- 246 Charbonnier, B., Laurent, C., Marchat, D.: Porous hydroxyapatite bioceramics produced by impregnation of 3D-printed wax mold: Slurry feature optimization, *J. Eur. Ceram. Soc.*, **36**, 4269–4279, (2016).
- 247 Roy, D.M., Linnehan, S.K.: Hydroxyapatite formed from coral skeletal carbonate by hydrothermal exchange, *Nature*, **247**, 220–222, (1974).
- 248 Zhang, X., Vecchio, K.S.: Conversion of natural marine skeletons as scaffolds for bone tissue engineering, *Front. Mater. Sci.*, **7**, 103–117, (2013).
- 249 Yang, Y., Yao, Q., Pu, X., Hou, Z., Zhang, Q.: Biphasic calcium phosphate macroporous scaffolds derived from oyster shells for bone tissue engineering, *Chem. Eng. J.*, **173**, 837–845, (2011).
- 250 Thanh, T.N.X., Maruta, M., Tsuru, K., Matsuya, S., Ishikawa, K.: Three-dimensional porous carbonate apatite with sufficient

- mechanical strength as a bone substitute material, *Adv. Mater. Res.*, **891–892**, 1559–1564, (2014).
- 251 Moroni, L., de Wijn, J.R., van Blitterswijk, C.A.: Integrating novel technologies to fabricate smart scaffolds, *J. Biomater. Sci. Polymer Edn.*, **19**, 543–572, (2008).
- 252 Studart, A.R., Gonzenbach, U.T., Tervoort, E., Gauckler, L.J.: Processing routes to macroporous ceramics: a review, *J. Am. Ceram. Soc.*, **89**, 1771–1789, (2006).
- 253 Hing, K., Annaz, B., Saeed, S., Revell, P., Buckland, T.: Microporosity enhances bioactivity of synthetic bone graft substitutes, *J. Mater. Sci. Mater. Med.*, **16**, 467–475, (2005).
- 254 Wang, Z., Sakakibara, T., Sudo, A., Kasai, Y.: Porosity of β -tricalcium phosphate affects the results of lumbar posterolateral fusion, *J. Spinal Disord. Tech.*, **26**, E40–E45, (2013).
- 255 Lan Levengood, S.K., Polak, S.J., Wheeler, M.B., Maki, A.J., Clark, S.G., Jamison, R.D., Wagoner Johnson, A.J.: Multiscale osteointegration as a new paradigm for the design of calcium phosphate scaffolds for bone regeneration, *Biomaterials*, **31**, 3552–3563, (2010).
- 256 Ruksudjarit, A., Pengpat, K., Rujijanagul, G., Tunkasiri, T.: The fabrication of nanoporous hydroxyapatite ceramics, *Adv. Mater. Res.*, **47–50**, 797–800, (2008).
- 257 Murugan, R., Ramakrishna, S., Rao, K.P.: Nanoporous hydroxyl-carbonate apatite scaffold made of natural bone, *Mater. Lett.*, **60**, 2844–2847, (2006).
- 258 Li, Y., Tjandra, W., Tam, K.C.: Synthesis and characterization of nanoporous hydroxyapatite using cationic surfactants as templates, *Mater. Res. Bull.*, **43**, 2318–2326, (2008).
- 259 El Asri, S., Laghzizil, A., Saoiabi, A., Alaoui, A., El Abassi, K., M'hamdi, R., Coradin, T.: A novel process for the fabrication of nanoporous apatites from moroccan phosphate rock, *Colloid Surf. A*, **350**, 73–78, (2009).
- 260 Ramli, R.A., Adnan, R., Bakar, M.A., Masudi, S.M.: Synthesis and characterisation of pure nanoporous hydroxyapatite, *J. Phys. Sci.*, **22**, 25–37, (2011).
- 261 LeGeros, R.Z.: Calcium phosphate-based osteoinductive materials, *Chem. Rev.*, **108**, 4742–4753, (2008).
- 262 Prokopiev, O., Sevostianov, I.: Dependence of the mechanical properties of sintered hydroxyapatite on the sintering temperature, *Mater. Sci. Eng. A*, **431**, 218–227, (2006).
- 263 Daculsi, G., Jegoux F., Layrolle, P.: The micro macroporous biphasic calcium phosphate concept for bone reconstruction and tissue engineering. In: *Advanced Biomaterials: Fundamentals, Processing and Applications*. Basu, B., Katti, D.S., Kumar, A. Eds. American Ceramic Society, Wiley, Hoboken, NJ, USA, 101–141, (2009).
- 264 Shipman, P., Foster, G., Schoeninger, M.: Burnt bones and teeth: an experimental study of color, morphology, crystal structure and shrinkage, *J. Archaeol. Sci.*, **11**, 307–325, (1984).
- 265 Rice, R.W.: *Porosity of ceramics*. Marcel Dekker, New York, NY, USA, 560, (1998).
- 266 Wang, H., Zhai, L., Li, Y., Shi, T.: Preparation of irregular mesoporous hydroxyapatite, *Mater. Res. Bull.*, **43**, 1607–1614, (2008).
- 267 Fan, J., Lei, J., Yu, C., Tu, B., Zhao, D.: Hard-templating synthesis of a novel rod-like nanoporous calcium phosphate bioceramics and their capacity as antibiotic carriers, *Mater. Chem. Phys.*, **103**, 489–493, (2007).
- 268 Sopyan, I., Mel, M., Ramesh, S., Khalid, K.A.: Porous hydroxyapatite for artificial bone applications, *Sci. Technol. Adv. Mater.*, **8**, 116–123, (2007).
- 269 Hsu, Y.H., Turner, I.G., Miles, A.W.: Fabrication of porous bioceramics with porosity gradients similar to the bimodal structure of cortical and cancellous bone, *J. Mater. Sci. Mater. Med.*, **18**, 2251–2256, (2007).
- 270 Abdurrahim, T., Sopyan, I.: Recent progress on the development of porous bioactive calcium phosphate for biomedical applications, *Recent Pat. Biomed. Eng.*, **1**, 213–229, (2008).
- 271 Munch, E., Franco, J., Deville, S., Hunger, P., Saiz, E., Tomasia, A.P.: Porous ceramic scaffolds with complex architectures, *JOM*, **60**, 54–59, (2008).
- 272 Ohji, T., Fukushima, M.: Macro-porous ceramics: Processing and properties. *Int. Mater. Rev.*, **57**, 115–131, (2012).
- 273 Naqshbandi, A.R., Sopyan, I., Gunawan: Development of porous calcium phosphate bioceramics for bone implant applications: a review, *Rec. Pat. Mater. Sci.*, **6**, 238–252, (2013).
- 274 Yan, X., Yu, C., Zhou, X., Tang, J., Zhao, D.: Highly ordered mesoporous bioactive glasses with superior in vitro bone-forming bioactivities, *Angew. Chem. Int. Ed. Engl.*, **43**, 5980–5984, (2004).
- 275 Izquierdo-Barba, I., Ruiz-González, L., Doadrio, J.C., González-Calbet, J.M., Vallet-Regí, M.: Tissue regeneration: a new property of mesoporous materials, *Solid State Sci.*, **7**, 983–989, (2005).
- 276 Cosijns, A., Vervaeke, C., Luyten, J., Mullens, S., Siepmann, F., van Hoorebeke, L., Masschaele, B., Cnudde, V., Remon, J.P.: Porous hydroxyapatite tablets as carriers for low-dosed drugs, *Eur. J. Pharm. Biopharm.*, **67**, 498–506, (2007).
- 277 Uchida, A., Shinto, Y., Araki, N., Ono, K.: Slow release of anticancer drugs from porous calcium hydroxyapatite ceramic, *J. Orthop. Res.*, **10**, 440–445, (1992).
- 278 Shinto, Y., Uchida, A., Korkusuz, F., Araki, N., Ono, K.: Calcium hydroxyapatite ceramic used as a delivery system for antibiotics, *J. Bone. Joint. Surg. Br.*, **74**, 600–604, (1992).
- 279 Martin, R.B., Chapman, M.W., Sharkey, N.A., Zissimos, S.L., Bay, B., Shors, E.C.: Bone ingrowth and mechanical properties of coralline hydroxyapatite 1 yr after implantation, *Biomaterials*, **14**, 341–348, (1993).
- 280 Kazakia, G.J., Nauman, E.A., Ebenstein, D.M., Halloran, B.P., Keaveny, T.M.: Effects of in vitro bone formation on the mechanical properties of a trabeculated hydroxyapatite bone substitute, *J. Biomed. Mater. Res. A*, **77A**, 688–699, (2006).
- 281 Hing, K.A., Best, S.M., Tanner, K.E., Bonfield, W., Revell, P.A.: Mediation of bone ingrowth in porous hydroxyapatite bone graft substitutes, *J. Biomed. Mater. Res. A*, **68A**, 187–200, (2004).
- 282 Drouet, C., Leriche, A., Hampshire, S., Kashani, M., Stamboulis, A., Iafisco, M., Tampieri, A.: Types of ceramics: Material class. In: *Advances in ceramic biomaterials: Materials, devices and challenges*. Palmero, P., De Barra, E., Cambier, F. (Eds.). Woodhead Publishing Series in Biomaterials. Woodhead Publishing, Cambridge, MA, USA., 500 p., 21–82, (2017).
- 283 Peppas, N.A., Langer, R.: New challenges in biomaterials, *Science*, **263**, 1715–1720, (1994).
- 284 Hench, L.L.: Biomaterials: a forecast for the future, *Biomaterials*, **19**, 1419–1423, (1998).
- 285 Barrère, F., Mahmood, T.A., de Groot, K., van Blitterswijk, C.A.: Advanced biomaterials for skeletal tissue regeneration: instructive and smart functions, *Mater. Sci. Eng. R*, **59**, 38–71, (2008).
- 286 Deligianni, D.D., Katsala, N.D., Koutsoukos, P.G., Missirlis, Y.F.: Effect of surface roughness of hydroxyapatite on human bone marrow cell adhesion, proliferation, differentiation and detachment strength, *Biomaterials*, **22**, 87–96, (2001).
- 287 Fini, M., Giardino, R., Borsari, V., Torricelli, P., Rimondini, L., Giavaresi, G., Aldini, N.N.: In vitro behaviour of osteoblasts cultured on orthopaedic biomaterials with different surface roughness, uncoated and fluorohydroxyapatite-coated, relative to the in vivo osteointegration rate, *Int. J. Artif. Organs*, **26**, 520–528, (2003).

- ²⁸⁸ Sato, M., Webster, T.J.: Designing orthopedic implant surfaces: harmonization of nanotopographical and chemical aspects, *Nanomedicine*, **1**, 351–354, (2006).
- ²⁸⁹ Li, X., van Blitterswijk, C.A., Feng, Q., Cui, F., Watari, F.: The effect of calcium phosphate microstructure on bone-related cells in vitro, *Biomaterials*, **29**, 3306–3316, (2008).
- ²⁹⁰ Kumar, G., Waters, M.S., Farooque, T.M., Young, M.F., Simon, C.G.: Freeform fabricated scaffolds with roughened struts that enhance both stem cell proliferation and differentiation by controlling cell shape, *Biomaterials*, **33**, 4022–4030, (2012).
- ²⁹¹ Holthaus, M.G., Treccani, L., Rezwan, K.: Osteoblast viability on hydroxyapatite with well-adjusted submicron and micron surface roughness as monitored by the proliferation reagent WST2–1, *J. Biomater. Appl.*, **27**, 791–800, (2013).
- ²⁹² Liu, H., Webster, T.J.: Nanomedicine for implants: a review of studies and necessary experimental tools, *Biomaterials*, **28**, 354–369, (2007).
- ²⁹³ Wang, C., Duan, Y., Markovic, B., Barbara, J., Howlett, C.R., Zhang, X., Zreiqat, H.: Proliferation and bone-related gene expression of osteoblasts grown on hydroxyapatite ceramics sintered at different temperature, *Biomaterials*, **25**, 2949–2956, (2004).
- ²⁹⁴ Samavedi, S., Whittington, A.R., Goldstein, A.S.: Calcium phosphate ceramics in bone tissue engineering: a review of properties and their influence on cell behavior, *Acta Biomater.*, **9**, 8037–8045, (2013).
- ²⁹⁵ Quarto, R., Mastrogiacomo, M., Cancedda, R., Kutepov, S.M., Mukhachev, V., Lavroukov, A., Kon, E., Marcacci, M.: Repair of large bone defects with the use of autologous bone marrow stromal cells, *N. Engl. J. Med.*, **344**, 385–386, (2001).
- ²⁹⁶ Vacanti, C.A., Bonassar, L.J., Vacanti, M.P., Shufflebarger, J.: Replacement of an avulsed phalanx with tissue-engineered bone, *N. Engl. J. Med.*, **344**, 1511–1514, (2001).
- ²⁹⁷ Morishita, T., Honoki, K., Ohgushi, H., Kotobuki, N., Matsushima, A., Takakura, Y.: Tissue engineering approach to the treatment of bone tumors: three cases of cultured bone grafts derived from patients' mesenchymal stem cells, *Artif. Organs*, **30**, 115–118, (2006).
- ²⁹⁸ Eniwumide, J.O., Yuan, H., Cartmell, S.H., Meijer, G.J., de Bruijn, J.D.: Ectopic bone formation in bone marrow stem cell seeded calcium phosphate scaffolds as compared to autograft and (cell seeded) allograft, *Eur. Cell Mater.*, **14**, 30–39, (2007).
- ²⁹⁹ Zhuang, Y., Gan, Y., Shi, D., Zhao, J., Tang, T., Dai, K.: A novel cytotherapy device for rapid screening, enriching and combining mesenchymal stem cells into a biomaterial for promoting bone regeneration, *Sci. Rep.*, **7**, 15463, (2017).
- ³⁰⁰ Zuolin, J., Hong, Q., Jiali, T.: Dental follicle cells combined with beta-tricalcium phosphate ceramic: a novel available therapeutic strategy to restore periodontal defects, *Med. Hypotheses*, **75**, 669–670, (2010).
- ³⁰¹ Ge, S., Zhao, N., Wang, L., Yu, M., Liu, H., Song, A., Huang, J., Wang, G., Yang, P.: Bone repair by periodontal ligament stem cell-seeded nanohydroxyapatite-chitosan scaffold, *Int. J. Nanomed.*, **7**, 5405–5414, (2012).
- ³⁰² Franch, J., Díaz-Bertrana, C., Lafuente, P., Fontecha, P., Durall, I.: Beta-tricalcium phosphate as a synthetic cancellous bone graft in veterinary orthopaedics: A retrospective study of 13 clinical cases, *Vet. Comp. Orthop. Traumatol.*, **19**, 196–204, (2006).
- ³⁰³ Vertenten, G., Gasthuys, F., Cornelissen, M., Schacht, E., Vlaminck, L.: Enhancing bone healing and regeneration: present and future perspectives in veterinary orthopaedics, *Vet. Comp. Orthop. Traumatol.*, **23**, 153–162, (2010).